



COMENIUS UNIVERSITY

Faculty of Medicine

Department of Medical Chemistry, Biochemistry and Clinical Biochemistry



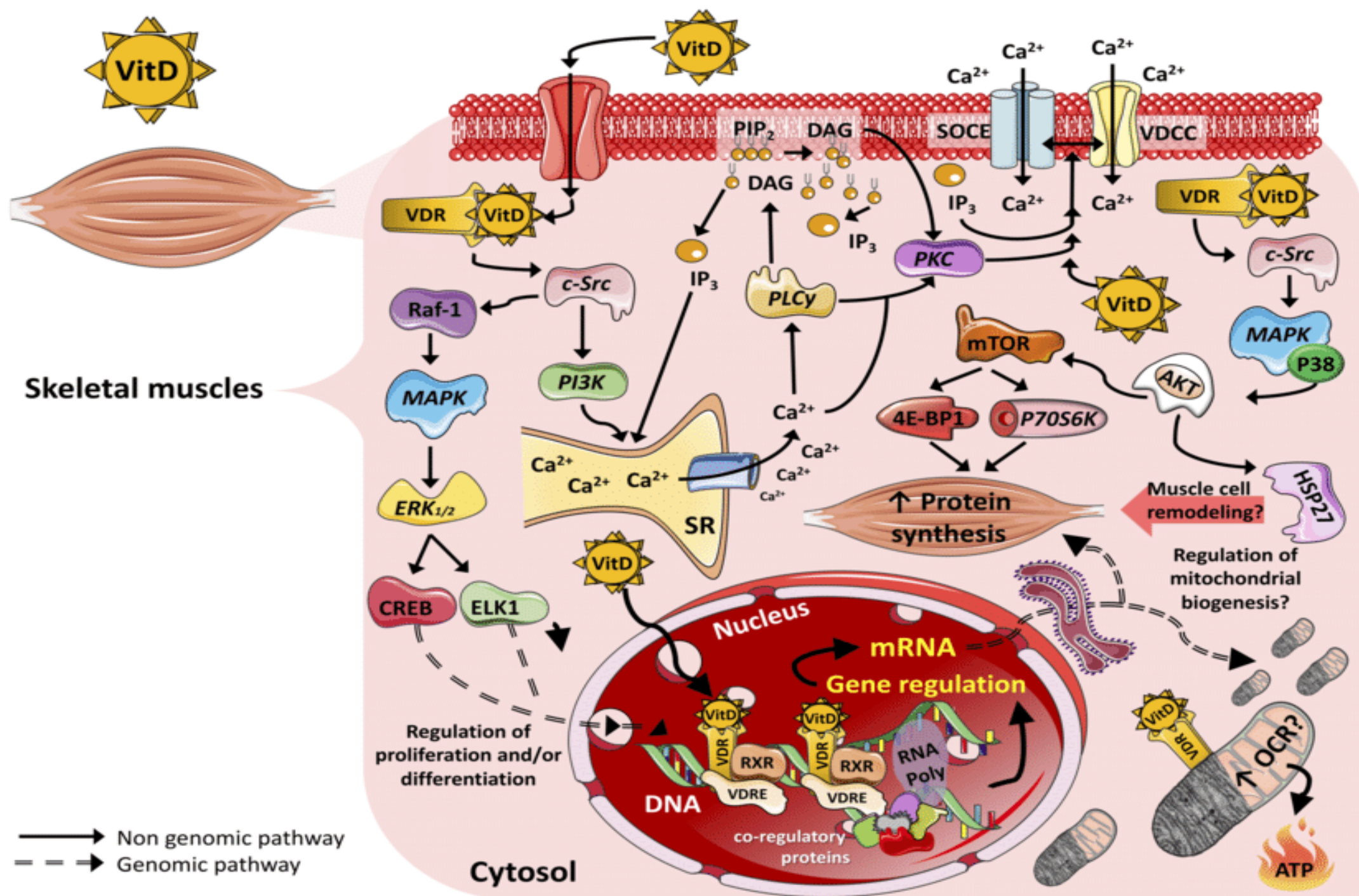
Effect of vitamin D on senescent cells

Ingrid Žitňanová, Mária Janubová, Petra Pazderová, Zuzana Sumbalová

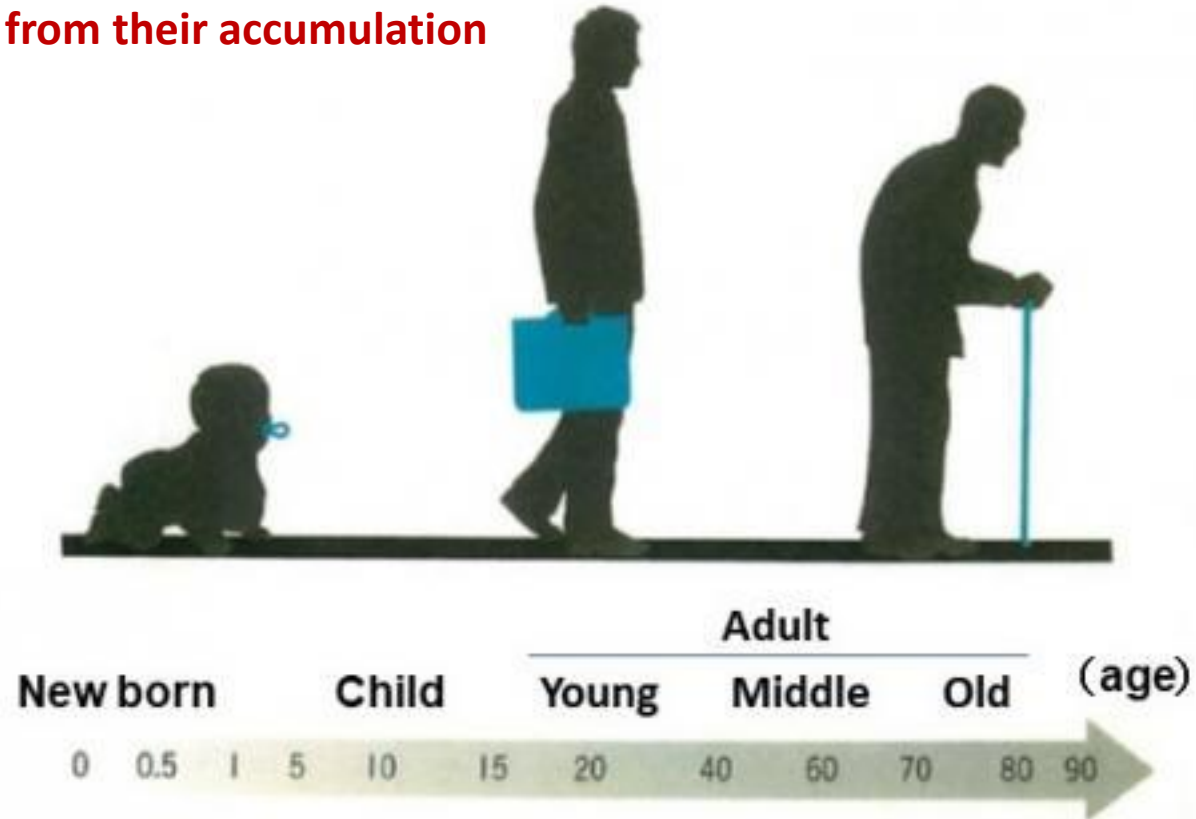
Nutriaging summer school

Ing. Katarína Koňariková, PhD.

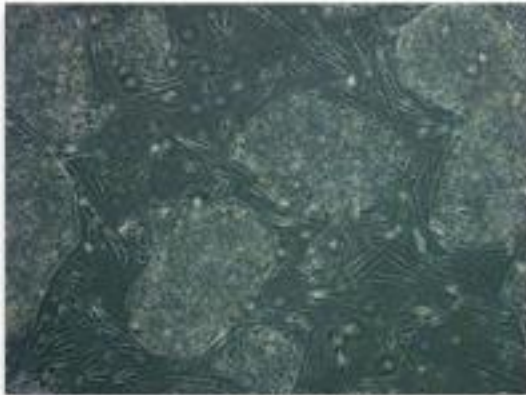
2022



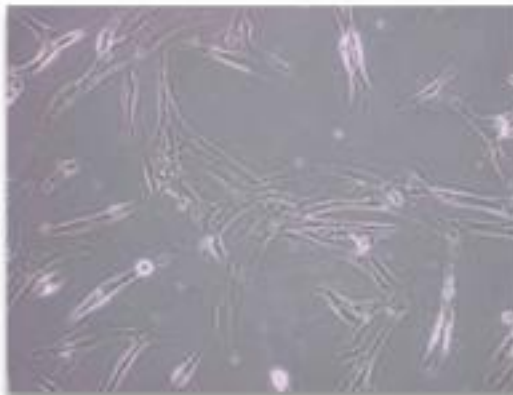
❑ The negative impacts results from their accumulation



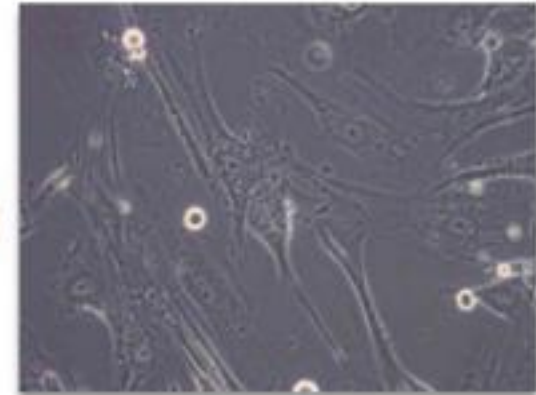
Stem cells



Differentiated cells



Senescent cells



Replicative model of senescence



We passaged cells up to the passage 21, when the senescence marker levels were the highest.

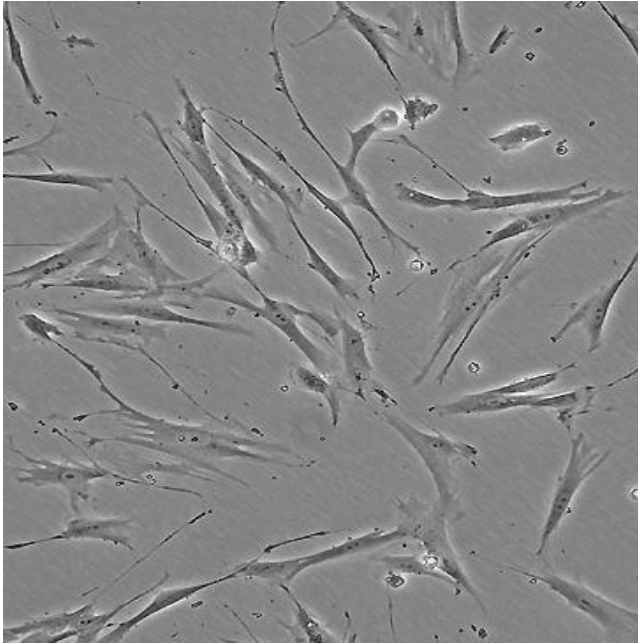
Stress induced senescence



Cells were treated with 100 μ M hydrogen peroxide for 30 min.

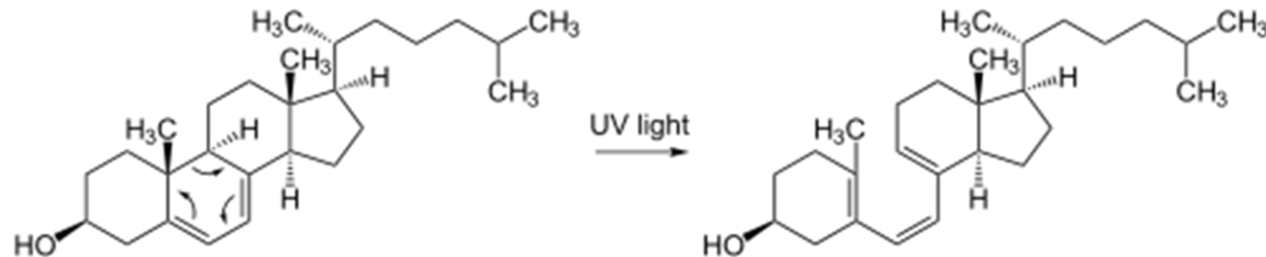
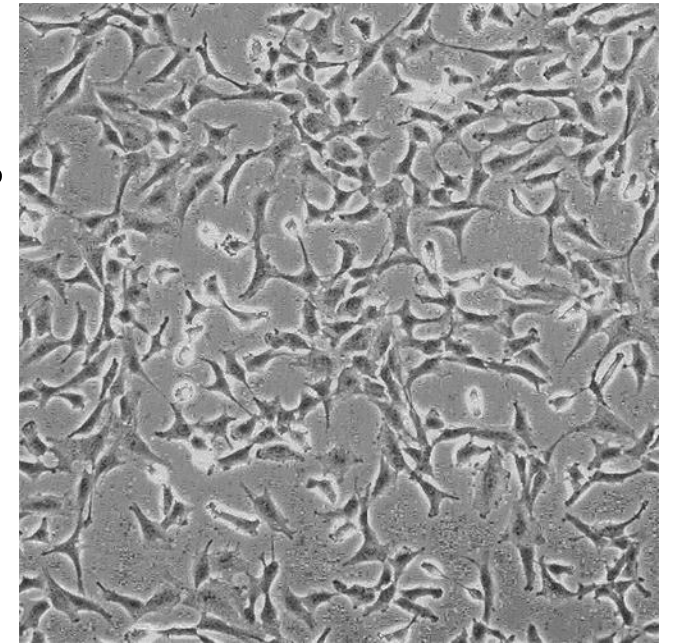
Materials and methods

Human lung cells (**MRC-5**)



- in passage 8-10 (p8-10) as normal cells
- in passage 21 (p21) as senescent cells
- senescence induced by 100 μM H_2O_2

Human astrocytes (**NHA5**)



Vitamin D

- 1, 10, 20, 25, 50, 100 nM
- in DMSO
- in EtOH

Methods	Analysis
MTT colorimetric technique	Viability of the cells
Muse [®] Cell Cycle Assay Kit	Distribution of cells in the individual phases of the cell cycle
Western blot analysis	Molecular mechanism
Quantitative Real-time PCR = qRT-PCR	Gene expression at the level of transcription
Mitochondrial respiration	Measurement of respiration

Methods

Analysis

MTT colorimetric technique

Viability of the cells

Muse® Cell Cycle Assay Kit

Distribution of cells in the individual phases of the cell cycle

Comet assay

Damage of DNA

Normoglycemic condition

Hyperglycemic condition

Western blot analysis

Molecular mechanism

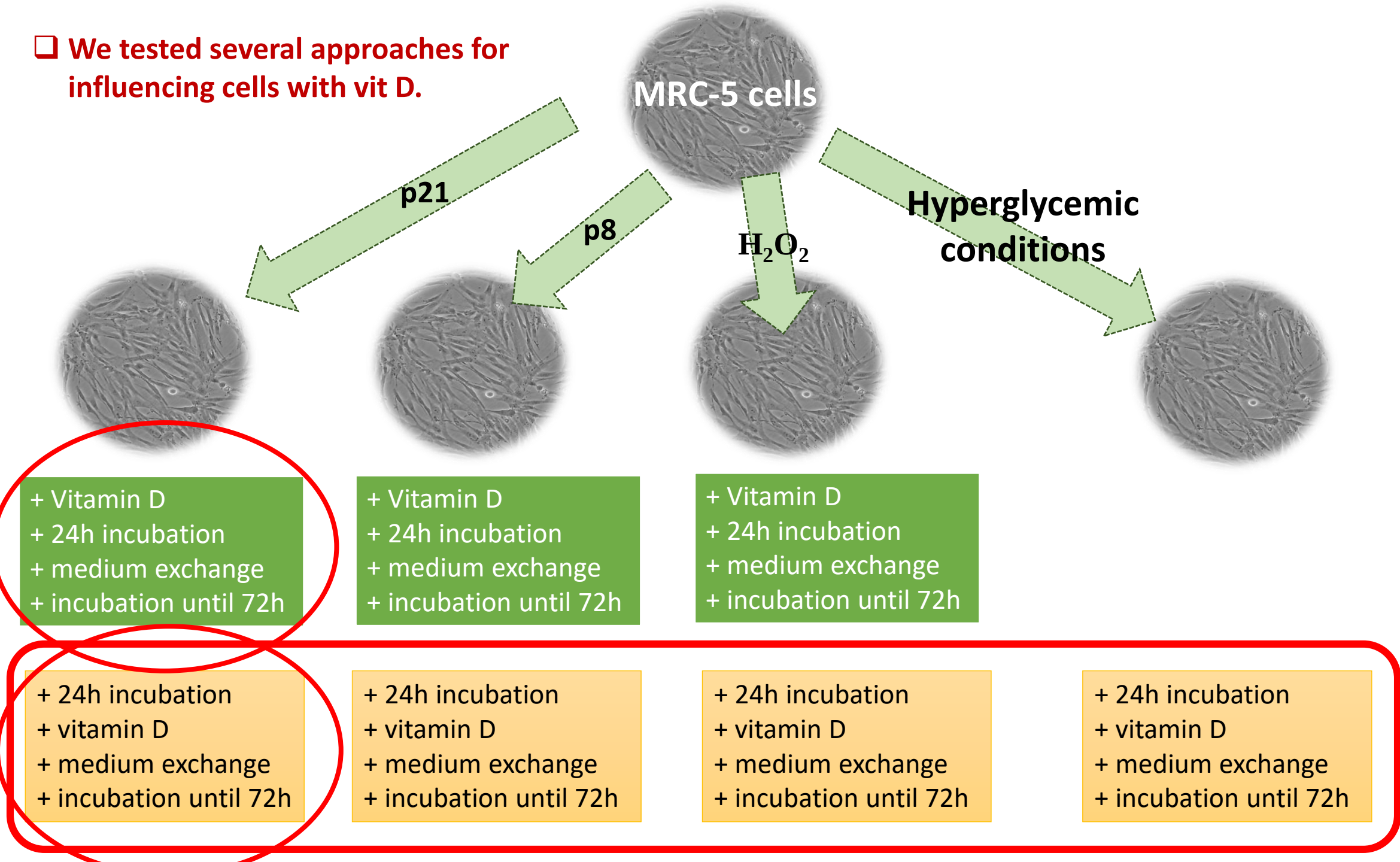
Quantitative Real-time PCR = qRT-PCR

Gene expression level at the level of transcription

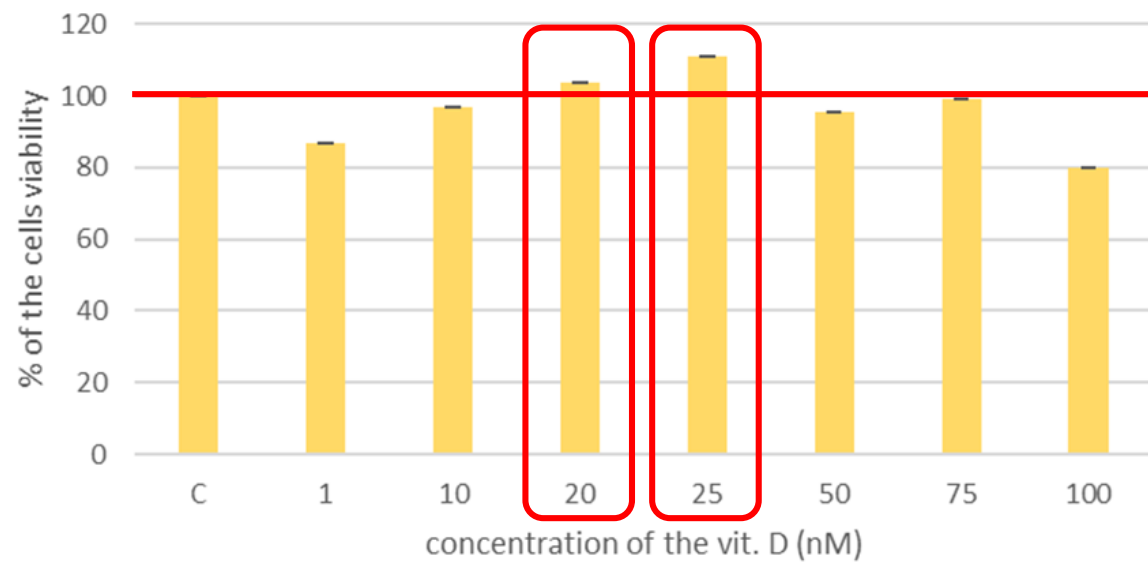
Mitochondrial respiration

Measurement of respiration

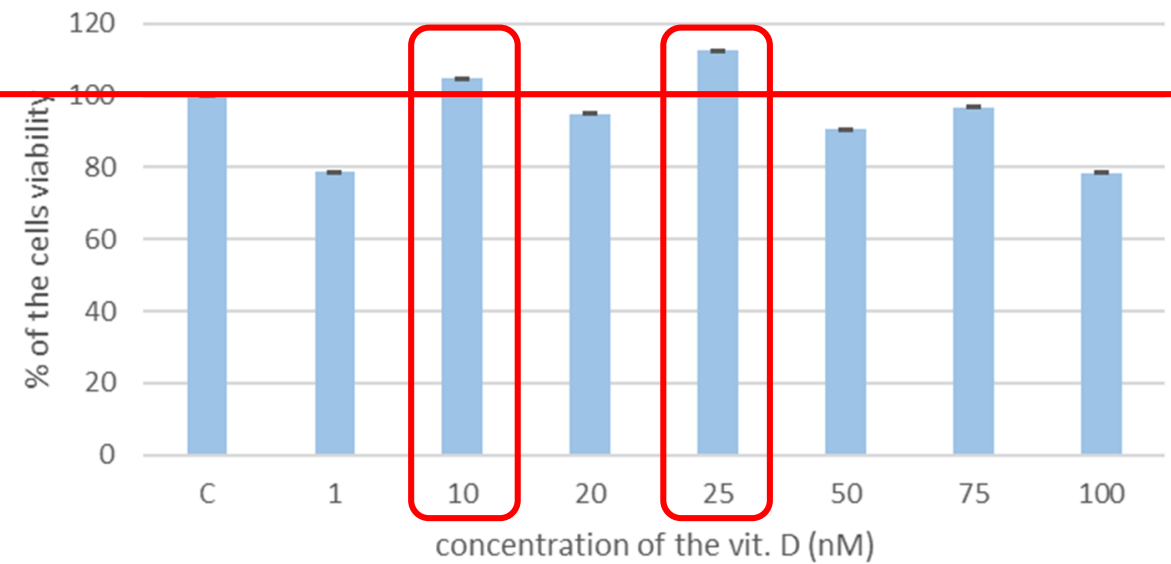
❑ We tested several approaches for influencing cells with vit D.



MRC-5, p8, 24h

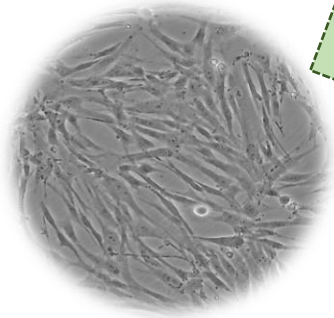


MRC-5, p21, 24h



MRC-5 cells

- + seeded the cells under HG conditions
- + incubated for 24 hours
- + affected cells with vitamin D for 24 hours



Vitamin D

Damaged the cells with H_2O_2 and then affected vitamin D

Added vitamin D to the cells and then damaged them with H_2O_2

not hyperglycemic
NG

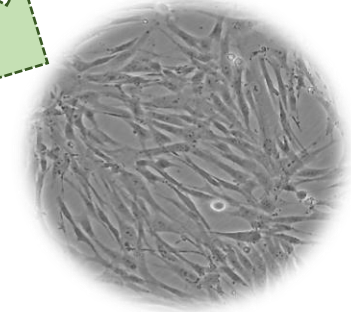


Vitamin D

Damaged the cells with H_2O_2 and then affected vitamin D

Added vitamin D to the cells and then damaged them with H_2O_2

- + seeded the cells under normal conditions
- + incubated for 24 h
- + changed the NG conditions to HG
- + addition of vitamin D

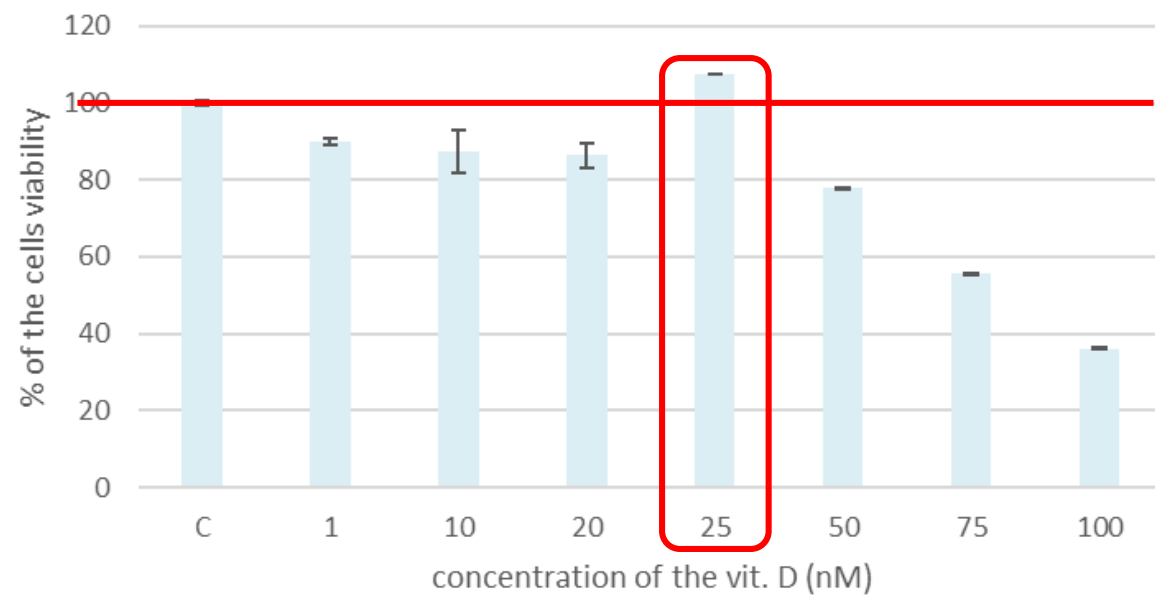


Vitamin D

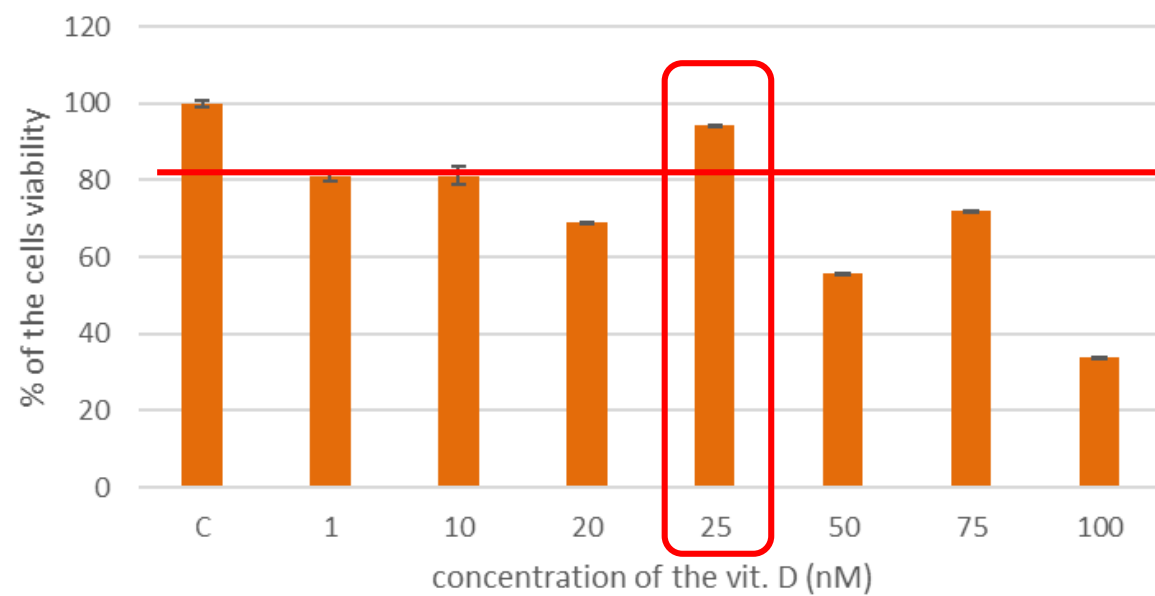
Damaged the cells with H_2O_2 and then affected vitamin D

Added vitamin D to the cells and then damaged them with H_2O_2

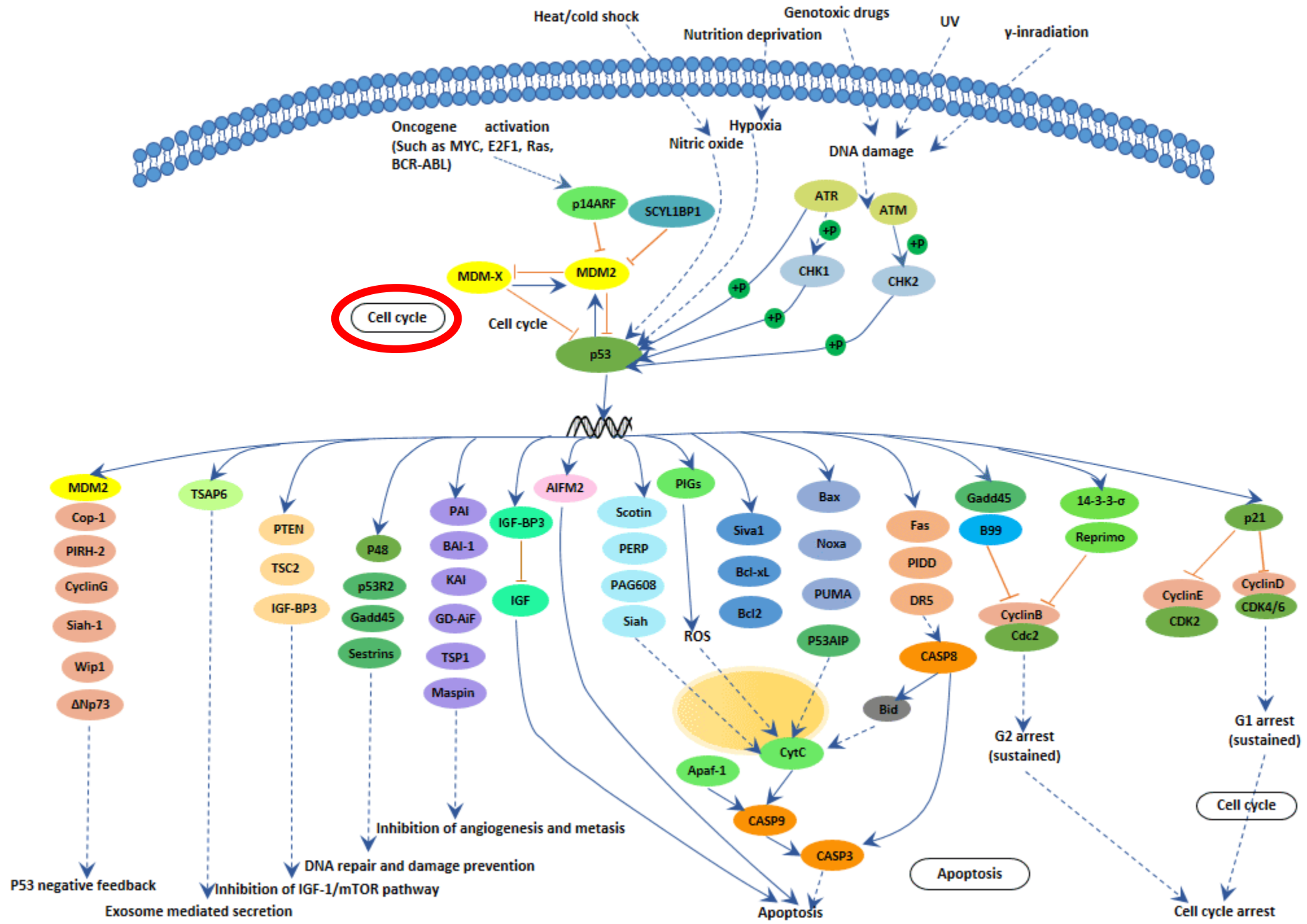
MRC-5, p10, 24h, HG



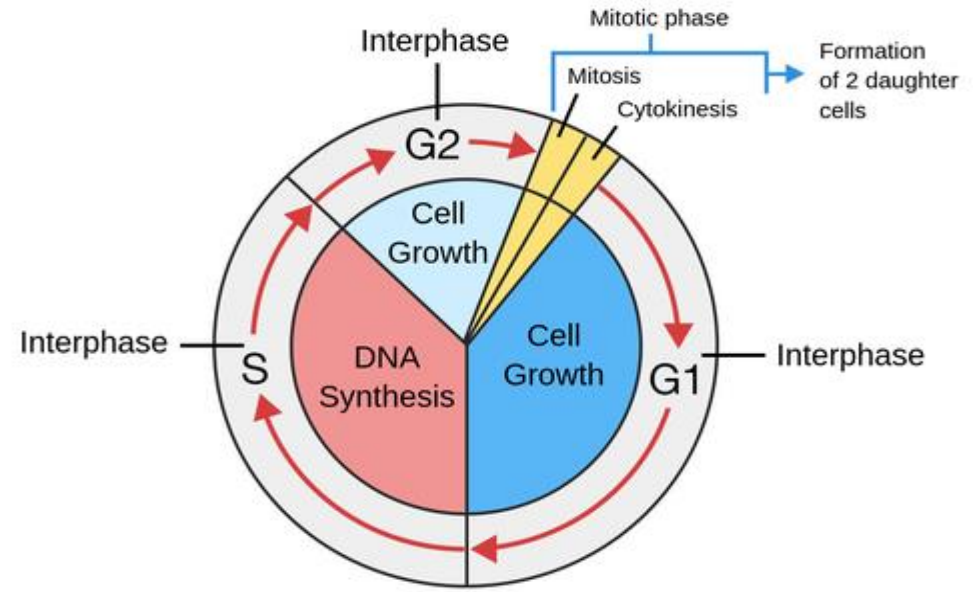
MRC-5, p21, 24h, HG



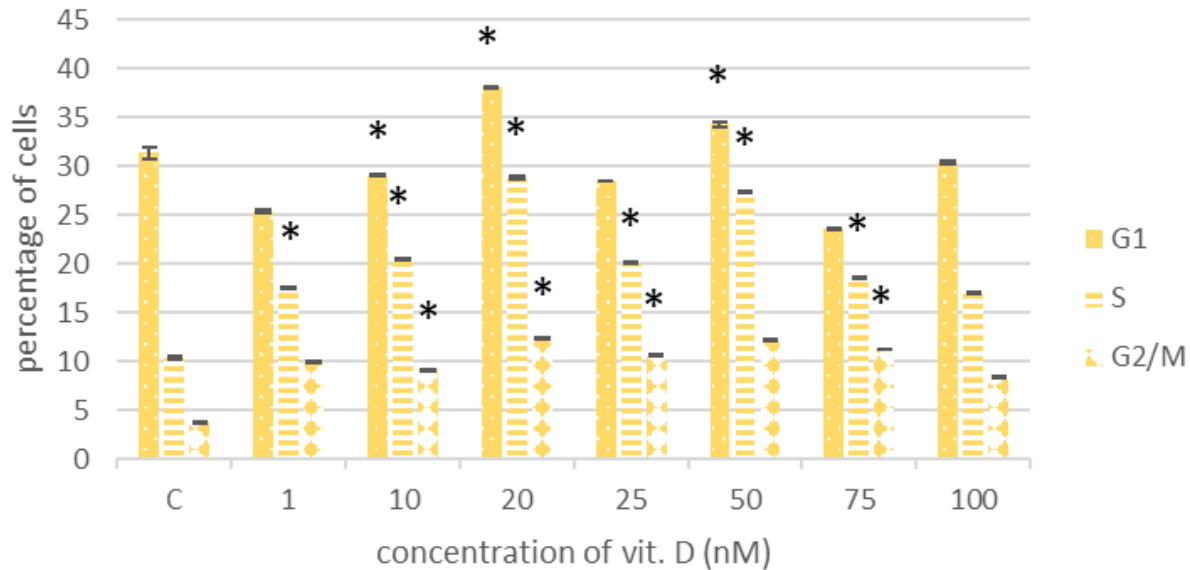
Methods	Analysis
MTT colorimetric technique	Viability of the cells
Muse [®] Cell Cycle Assay Kit	Distribution of cells in the individual phases of the cell cycle
Western blot analysis	Molecular mechanism
Quantitative Real-time PCR = qRT-PCR	Gene expression at the level of transcription
Mitochondrial respiration	Measurement of respiration



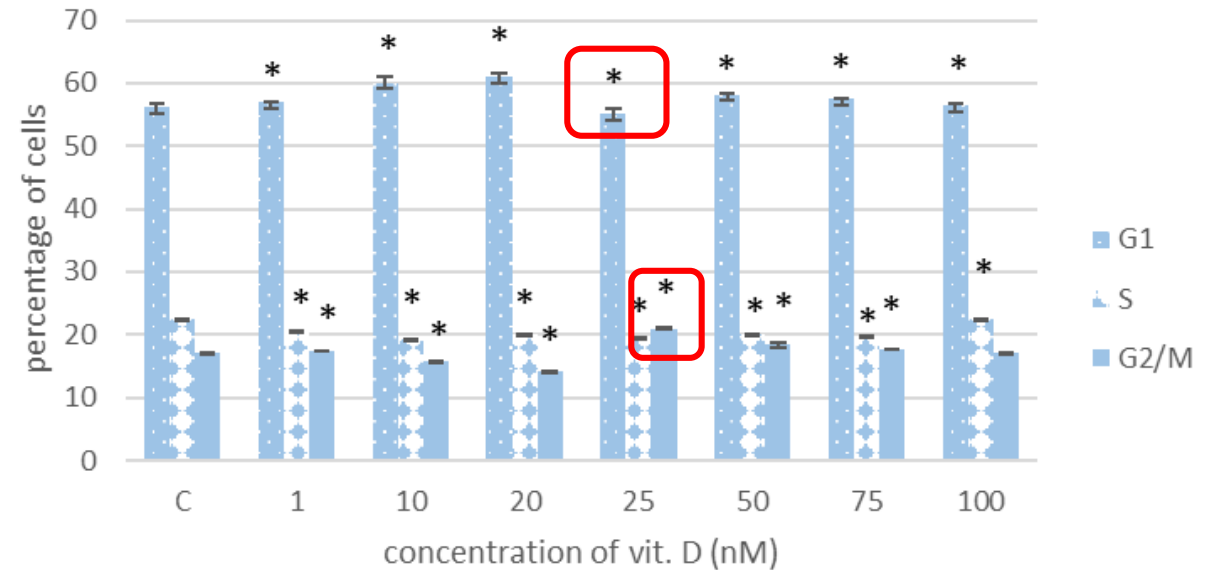
- **Muse® Cell Cycle Assay Kit**
- **Normoglycemic condition**

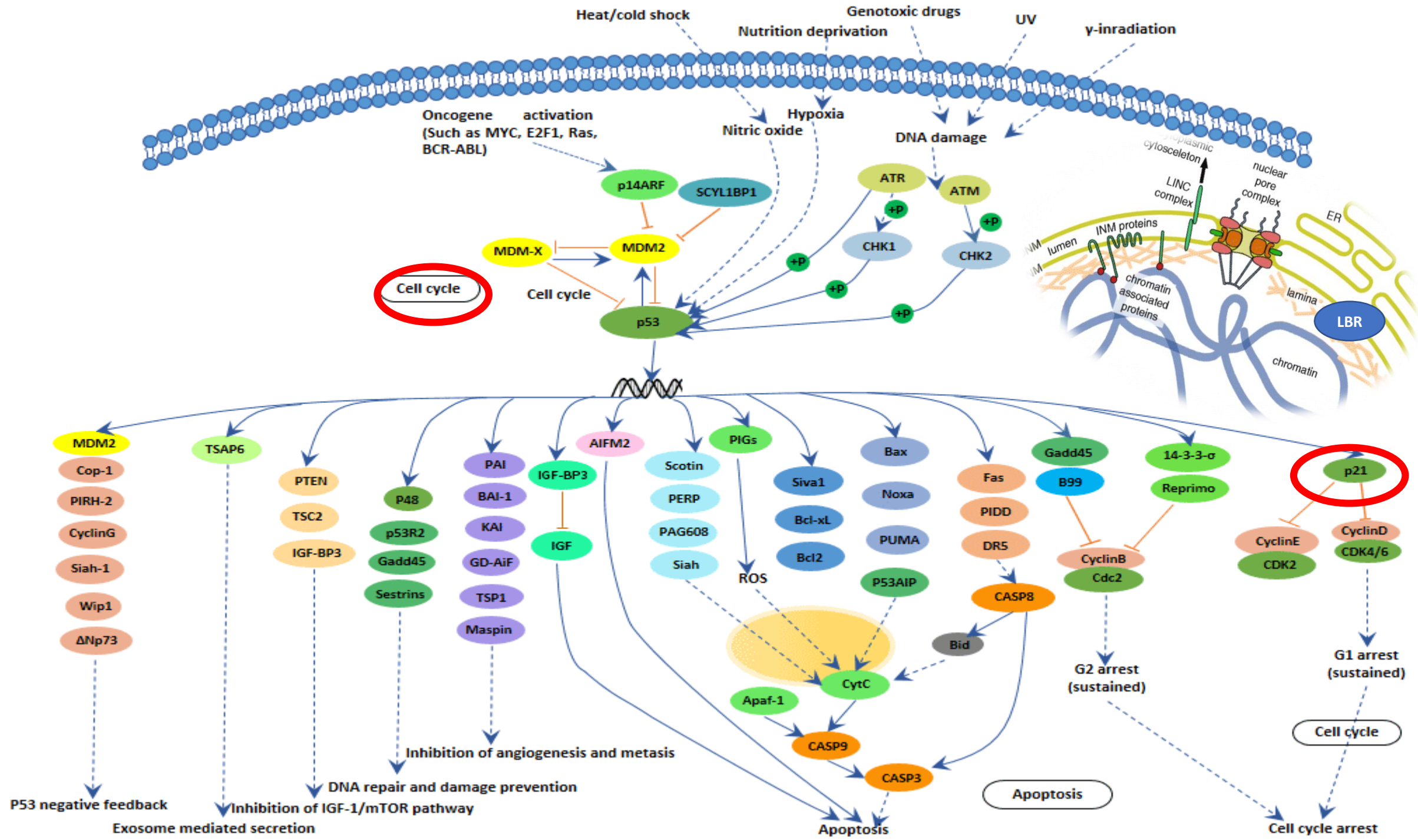


MRC-5, p8, 24h



MRC-5, p21, 24h





Methods	Analysis
MTT colorimetric technique	Viability of the cells
Muse [®] Cell Cycle Assay Kit	Distribution of cells in the individual phases of the cell cycle
Western blot analysis	Molecular mechanism
Quantitative Real-time PCR = qRT-PCR	Gene expression at the level of transcription
Mitochondrial respiration	Measurement of respiration

Programmed Cell Death

Autophagy

- Membrane blebbing
- Autophagic vacuoles
- Increased lysosomal activity

Programmed Necrosis

Pyroptosis

- Maintained mitochondria integrity
- Cell swelling
- Membrane rupture

Necroptosis

- Mitochondrial swelling
- Cell swelling
- Membrane rupture

Parthanatos

- Chromatine condensation
- Nuclear fragmentation

Ferroptosis

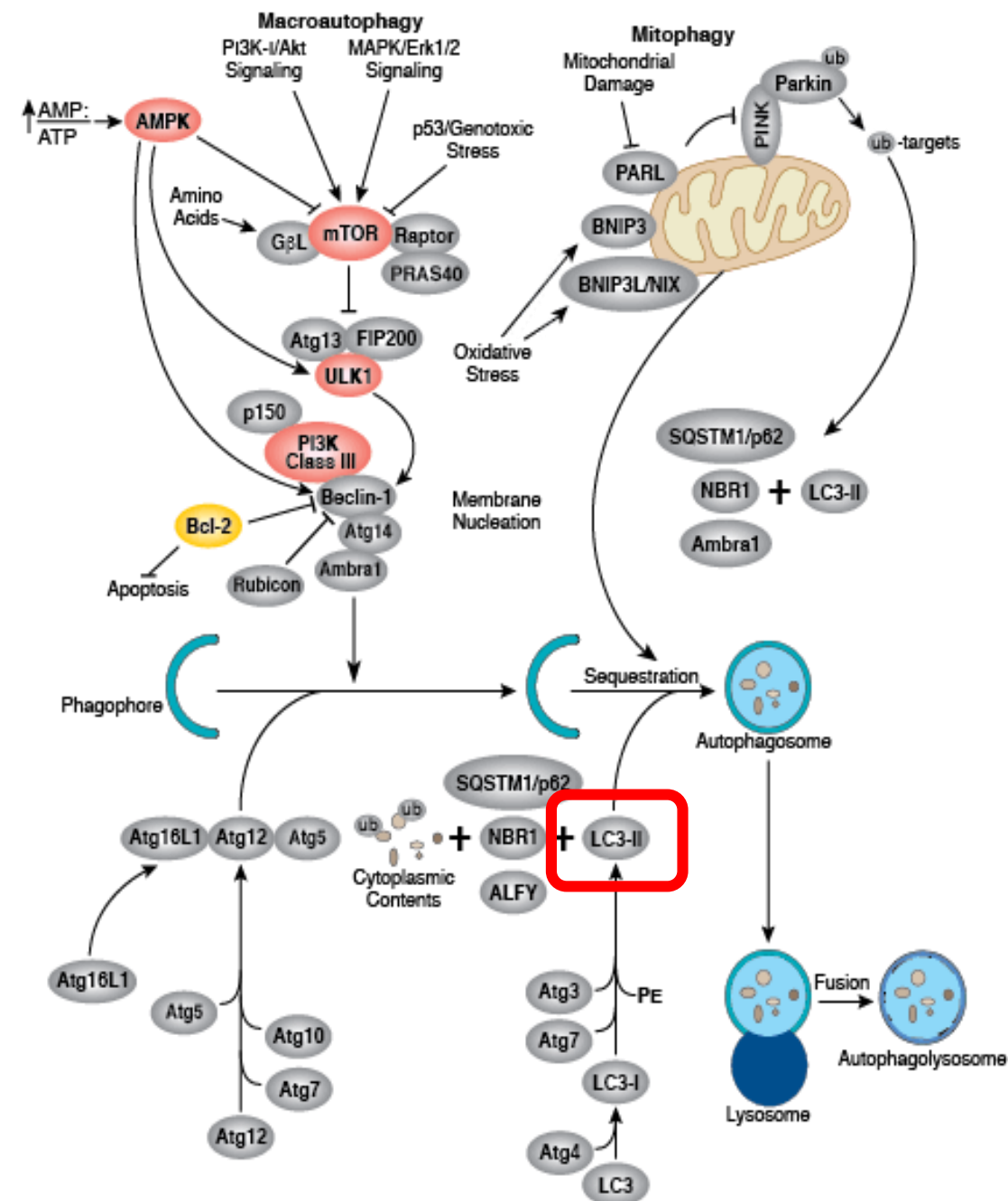
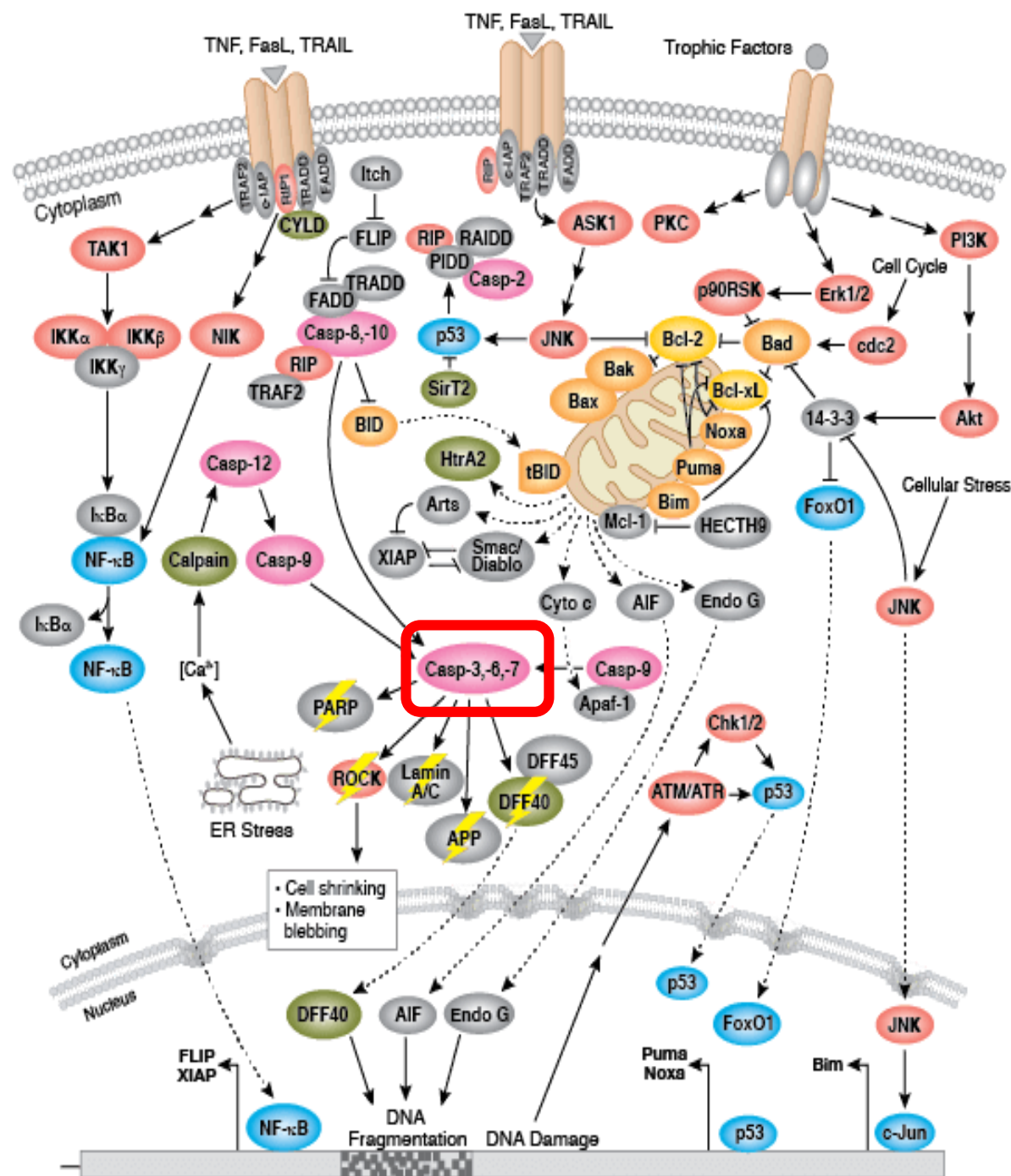
- Increased membrane density
- Small mitochondria

Apoptosis

- Chromatine condensation
- Nuclear fragmentation
- Apoptotic body
- Membrane Blebbing
- Cell shrinkage (in animal cells)

Non- Programmed Cell Death (Necrosis)

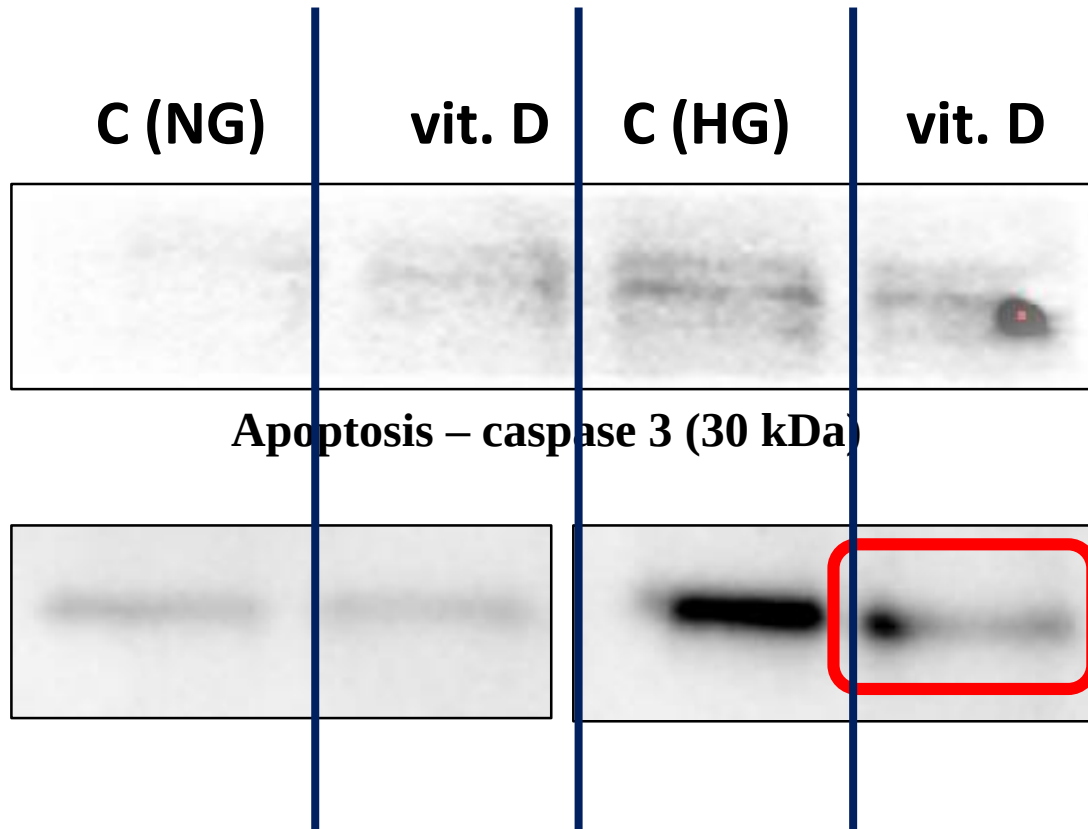
- Mitochondrial swelling
- Cell swelling
- Membrane rupture



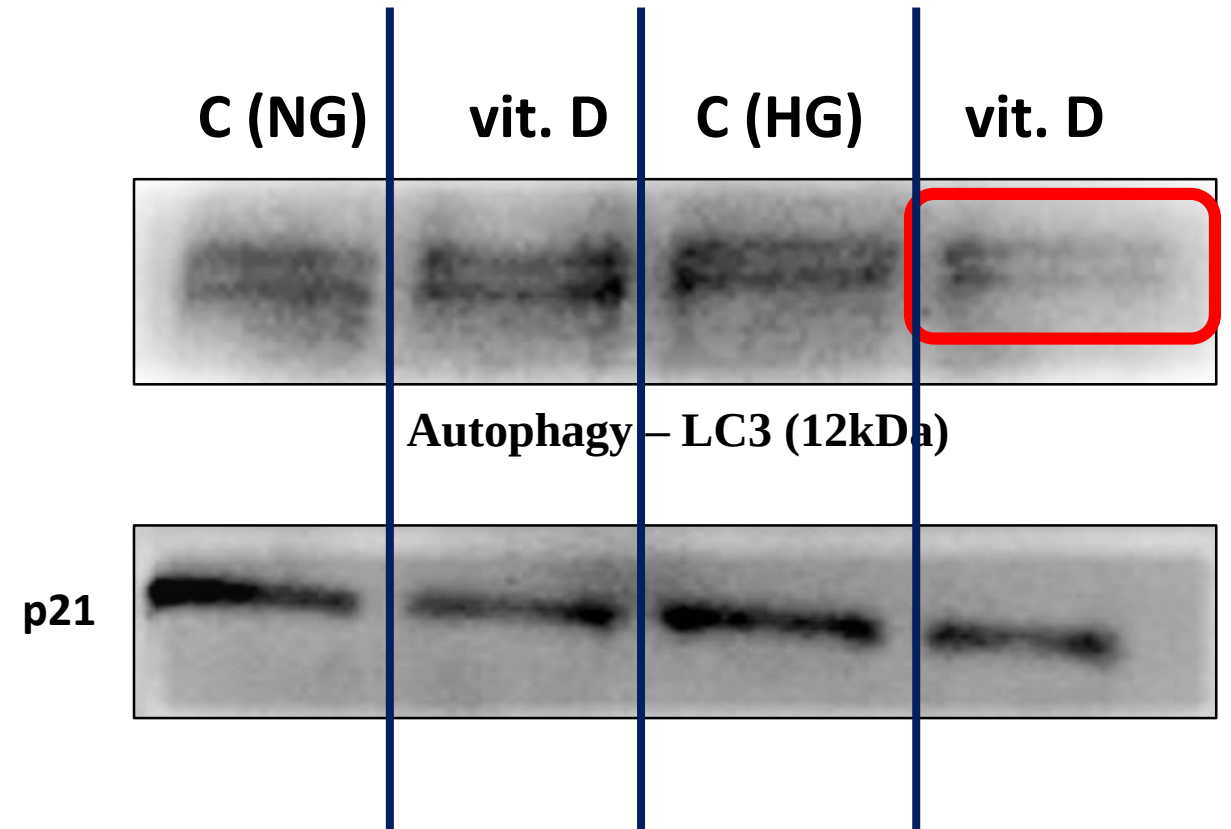
■ Western blot analysis

■ normo and hyperglycemic condition

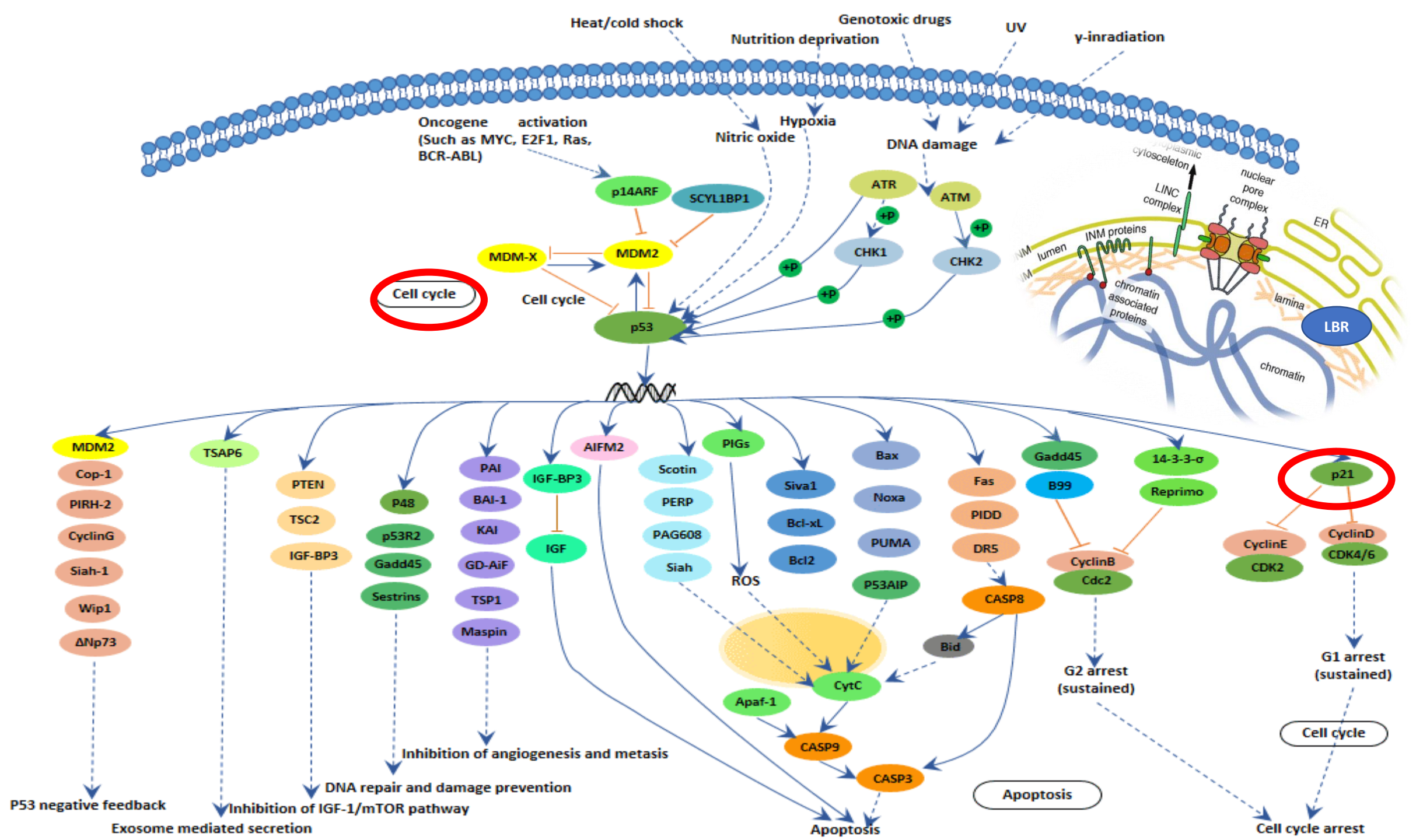
■ in passage 8 (p8) as normal cells



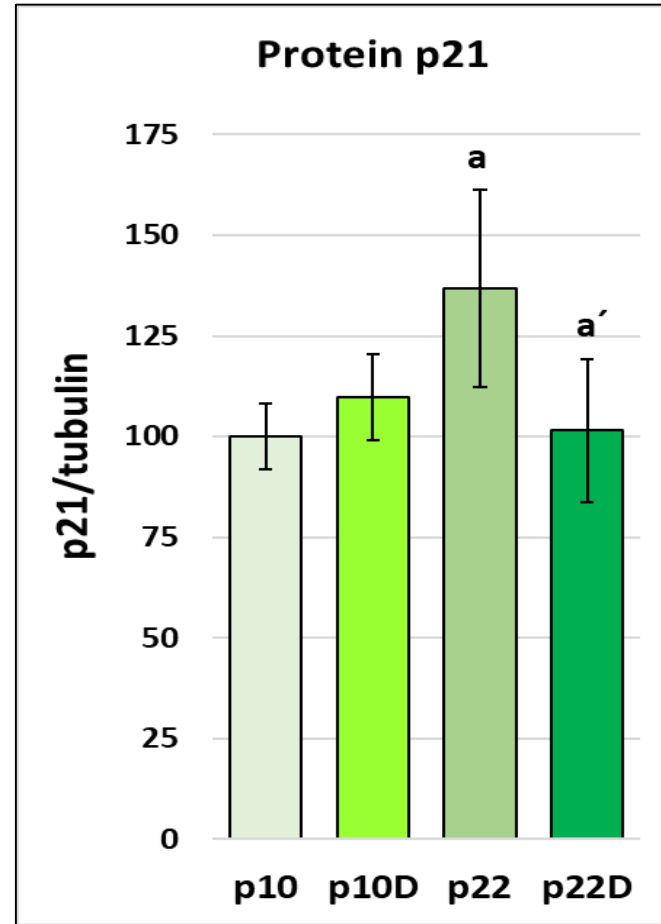
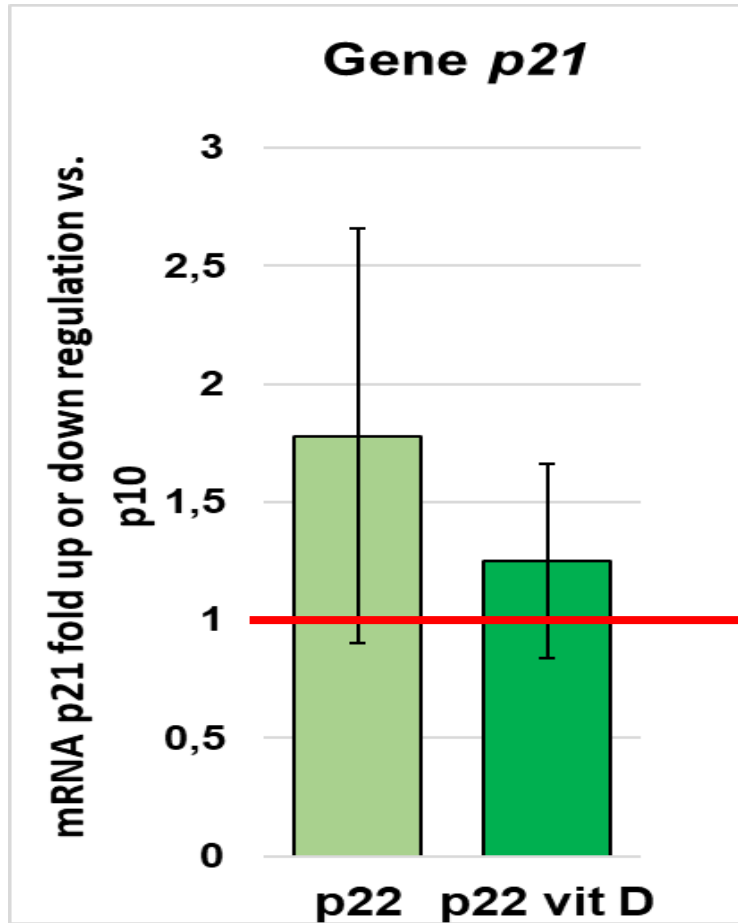
■ in passage 21 (p21) as senescent cells

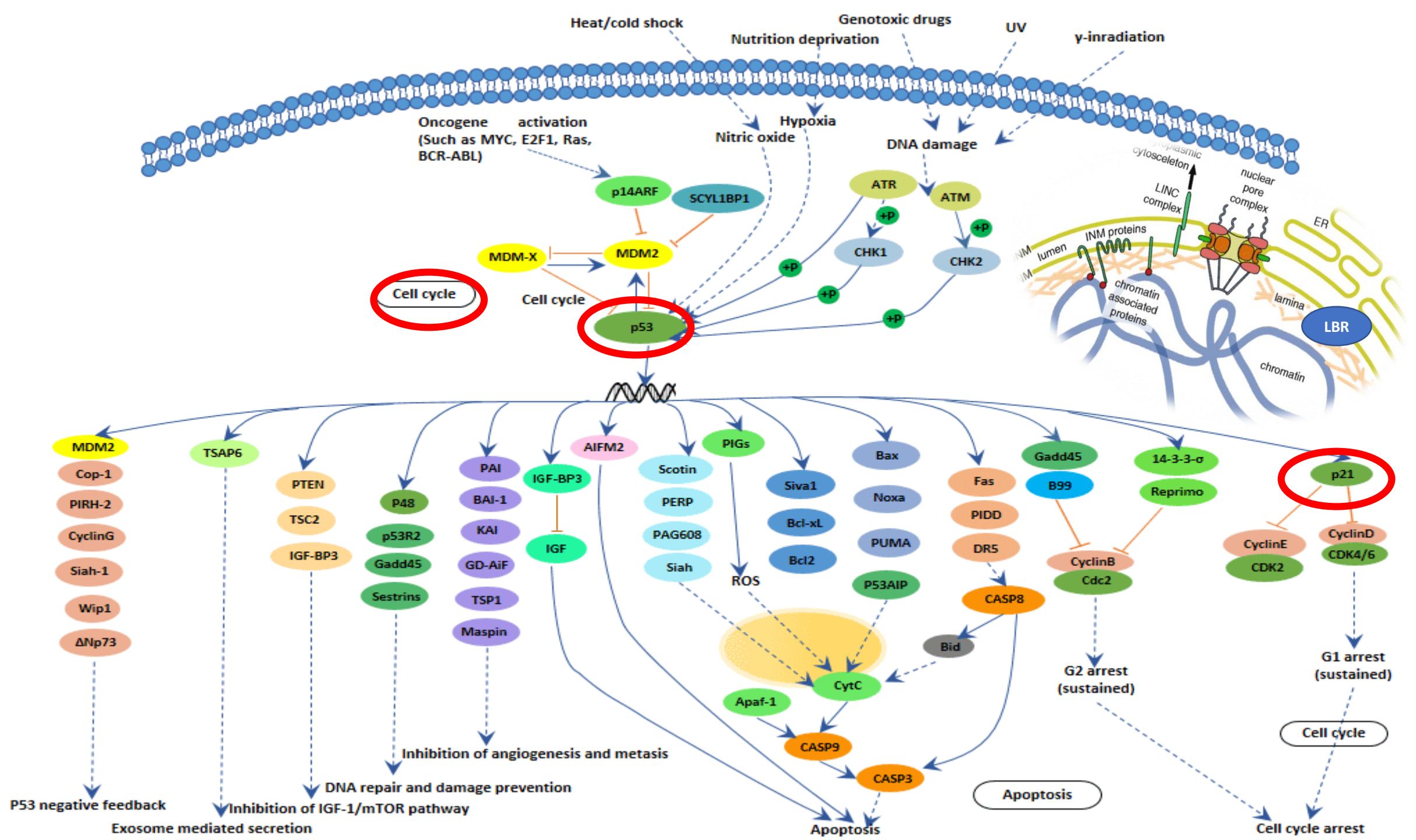


Methods	Analysis
MTT colorimetric technique	Viability of the cells
Muse [®] Cell Cycle Assay Kit	Distribution of cells in the individual phases of the cell cycle
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Quantitative Real-time PCR = qRT-PCR	Gene expression at the level of transcription
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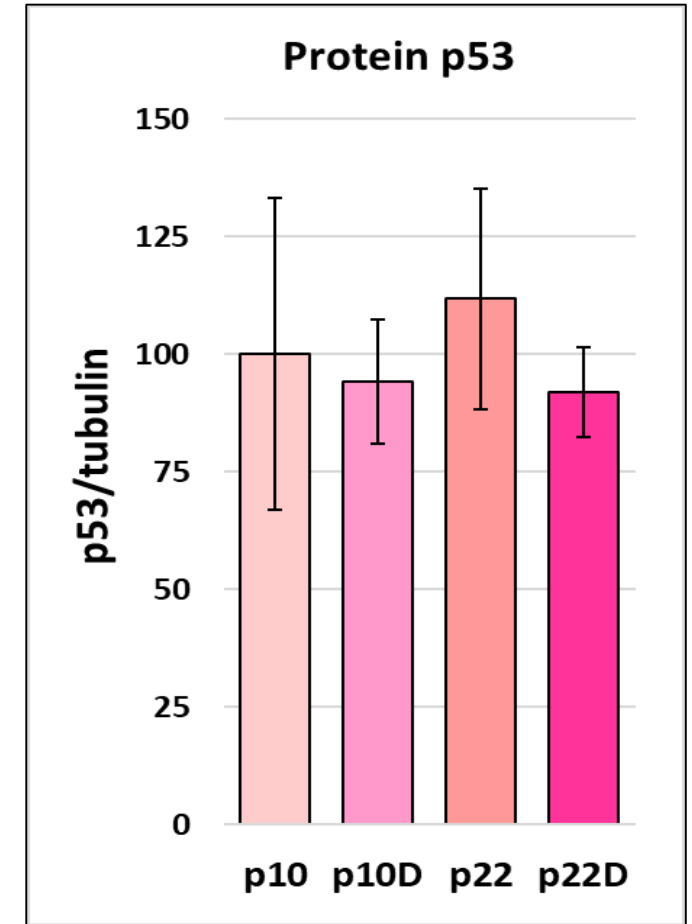
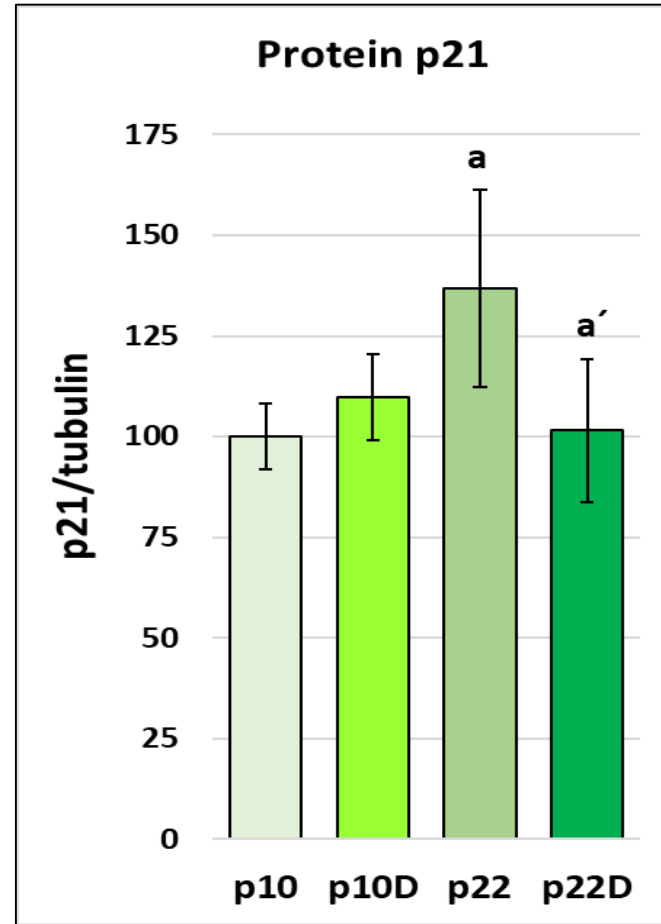
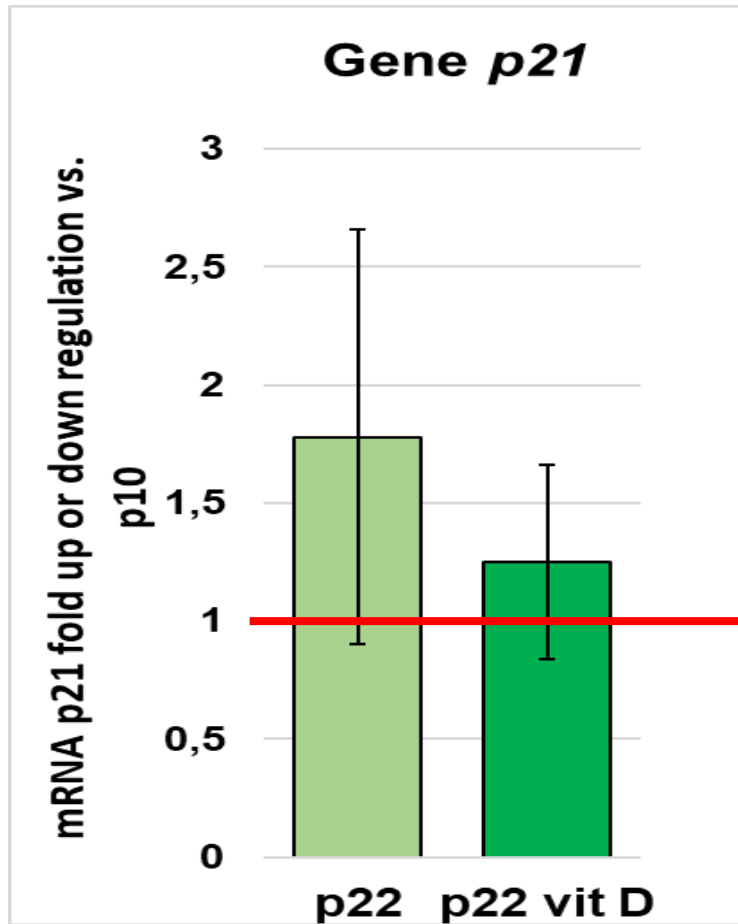


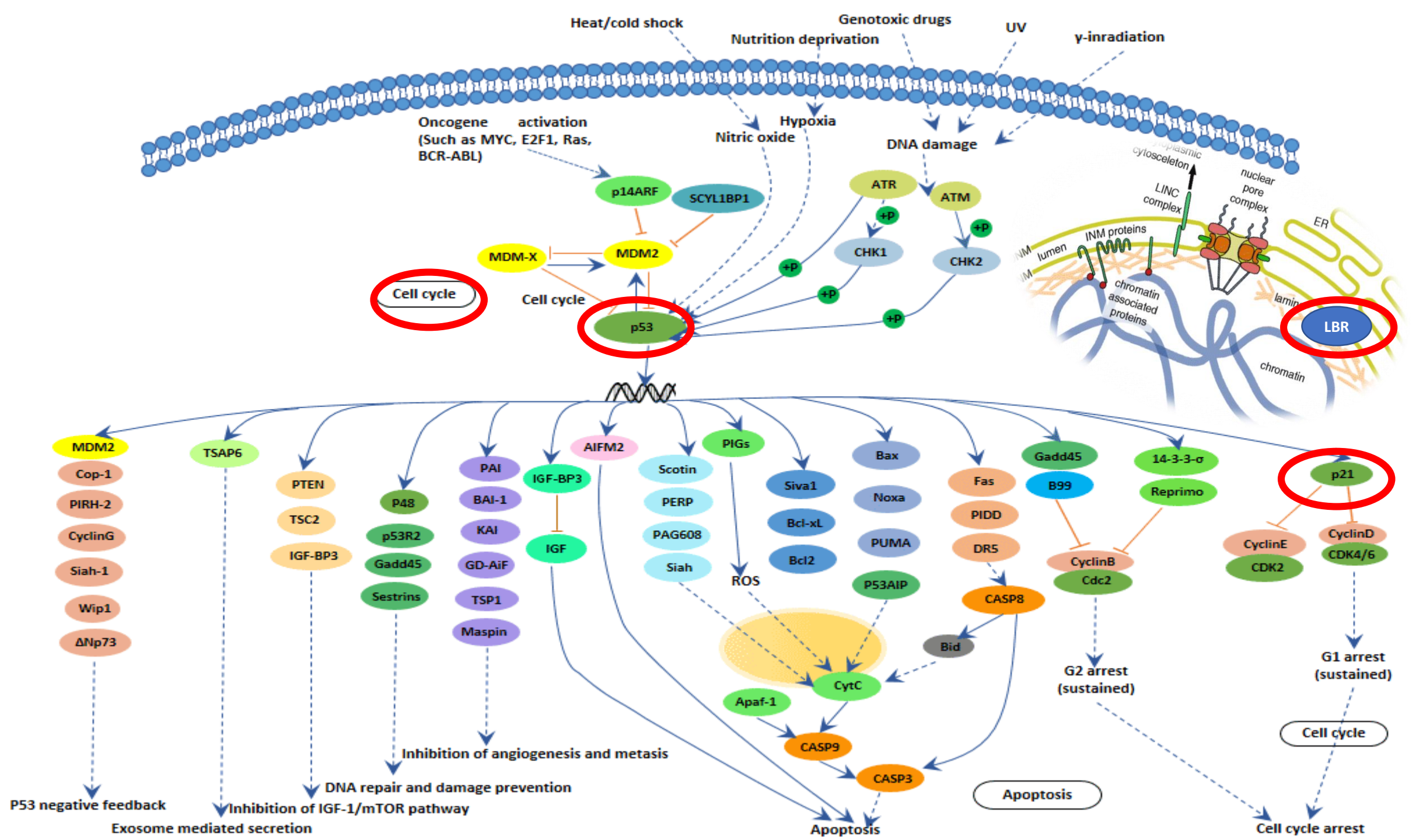
- quantitative Real-time PCR = qRT-PCR
 - Normoglycemic condition

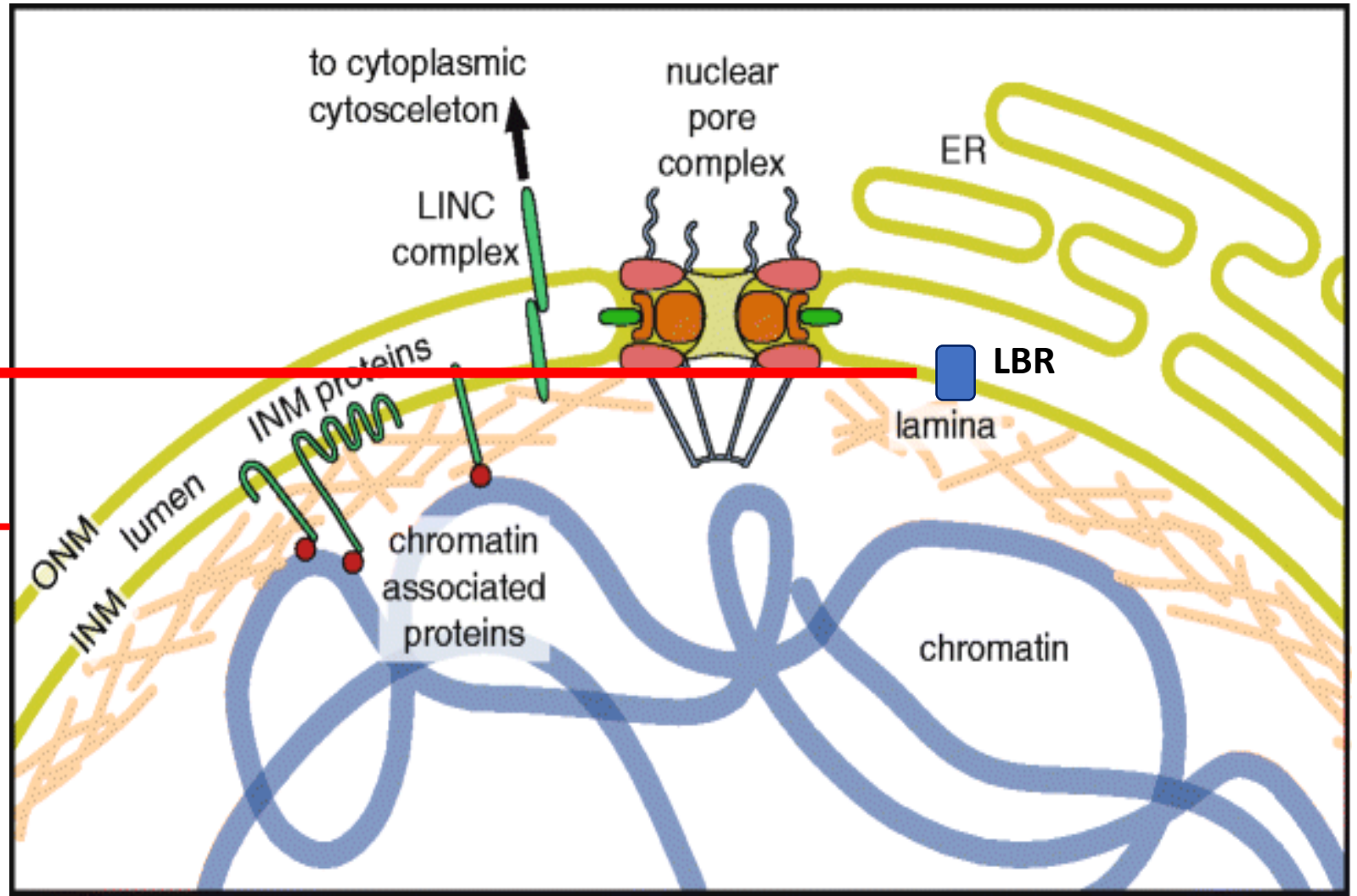
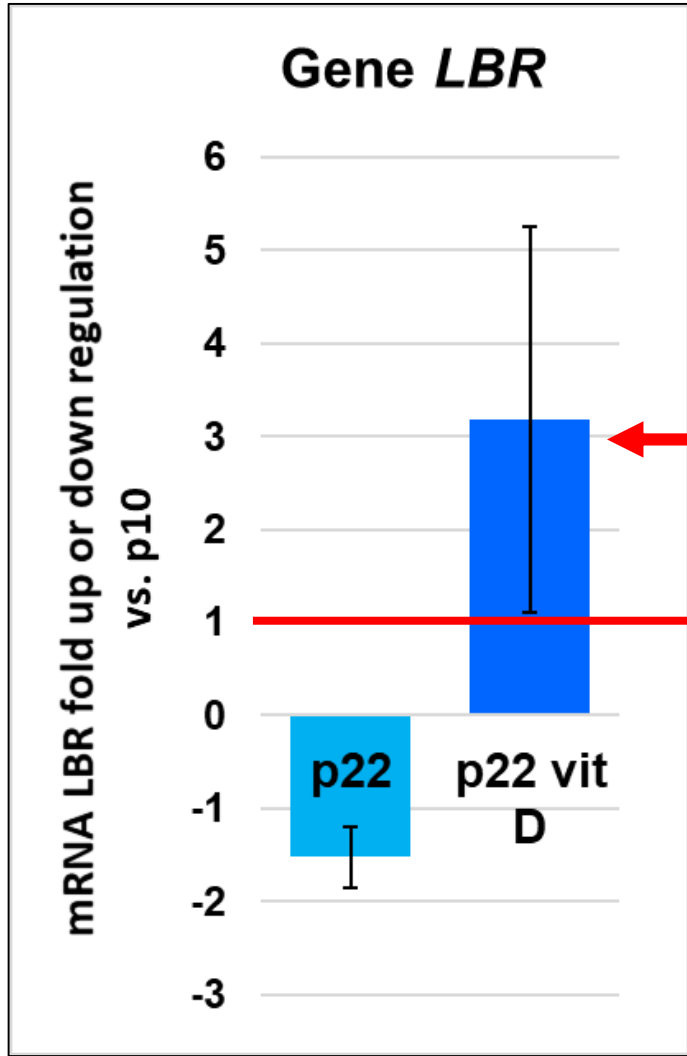




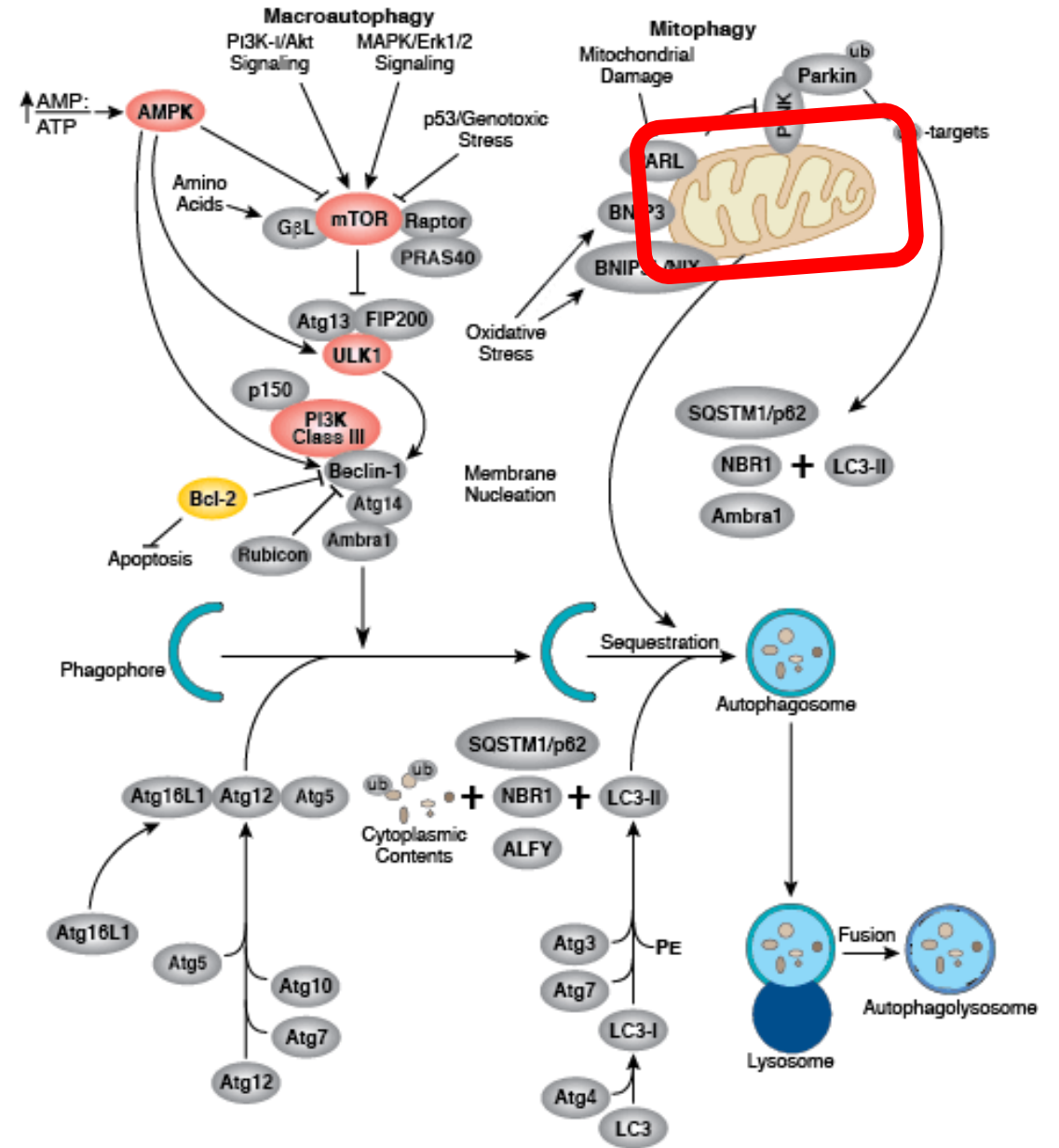
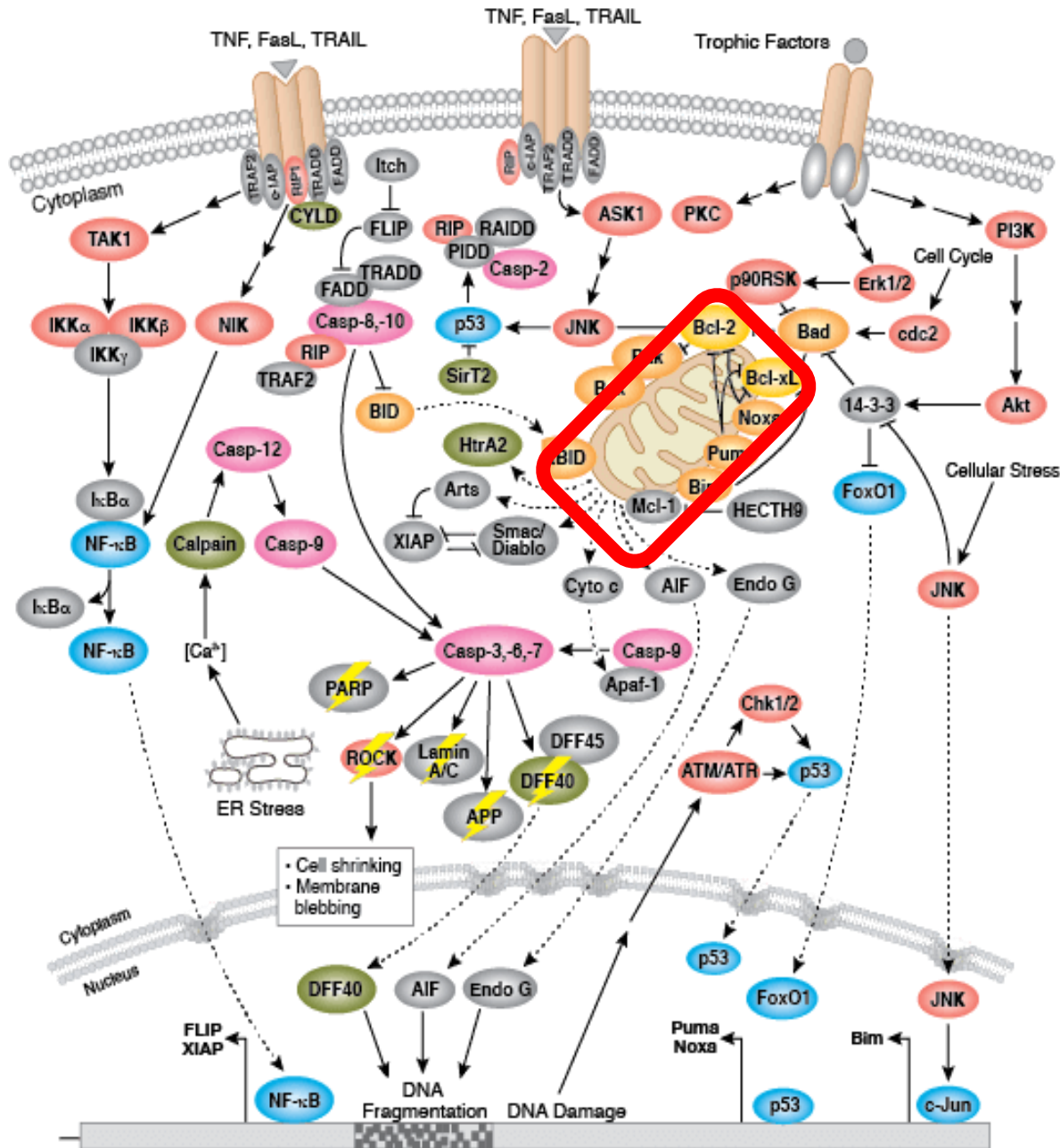
- quantitative Real-time PCR = qRT-PCR
 - Normoglycemic condition





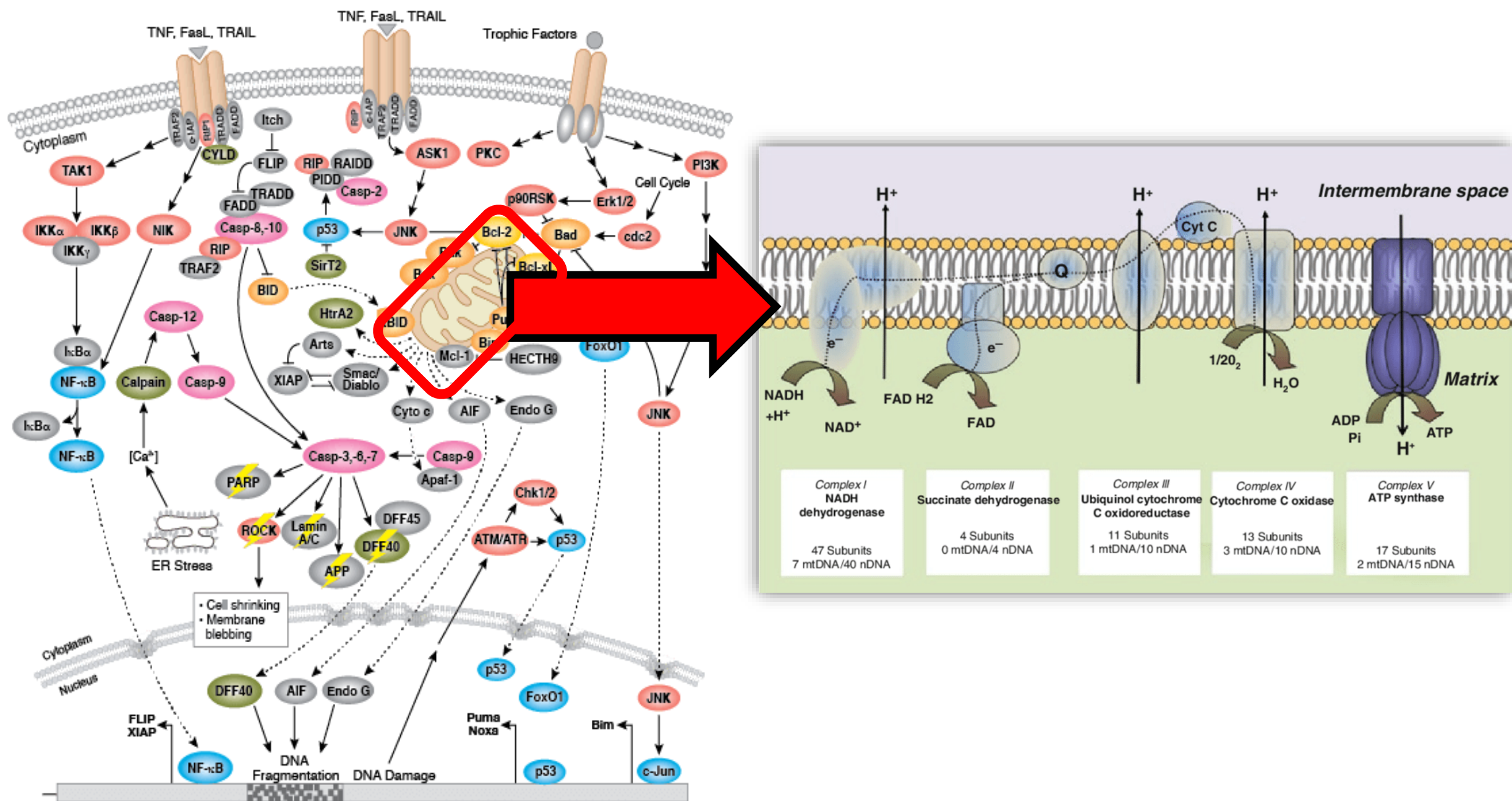


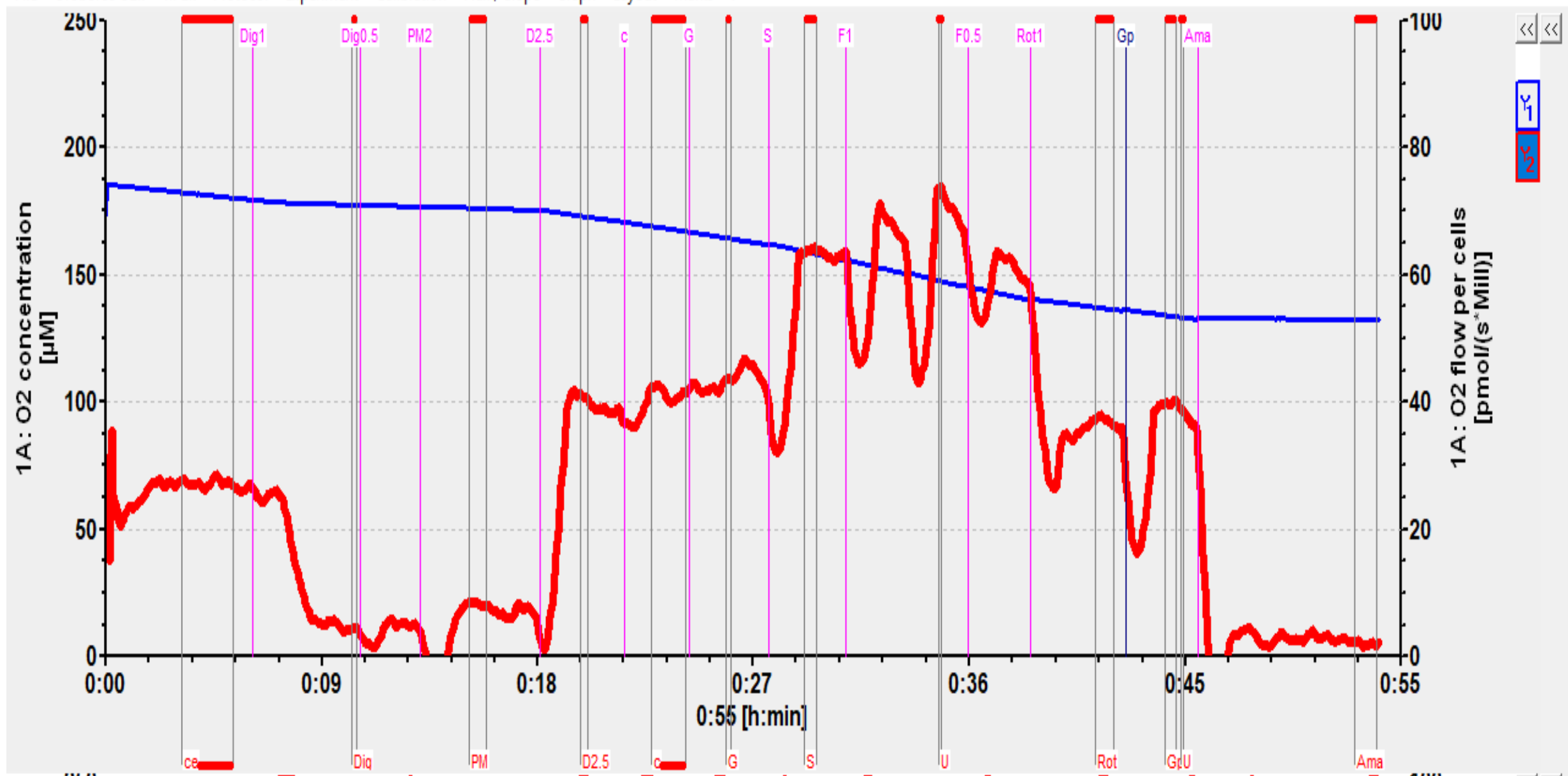
- **Mitochondrial respiration**

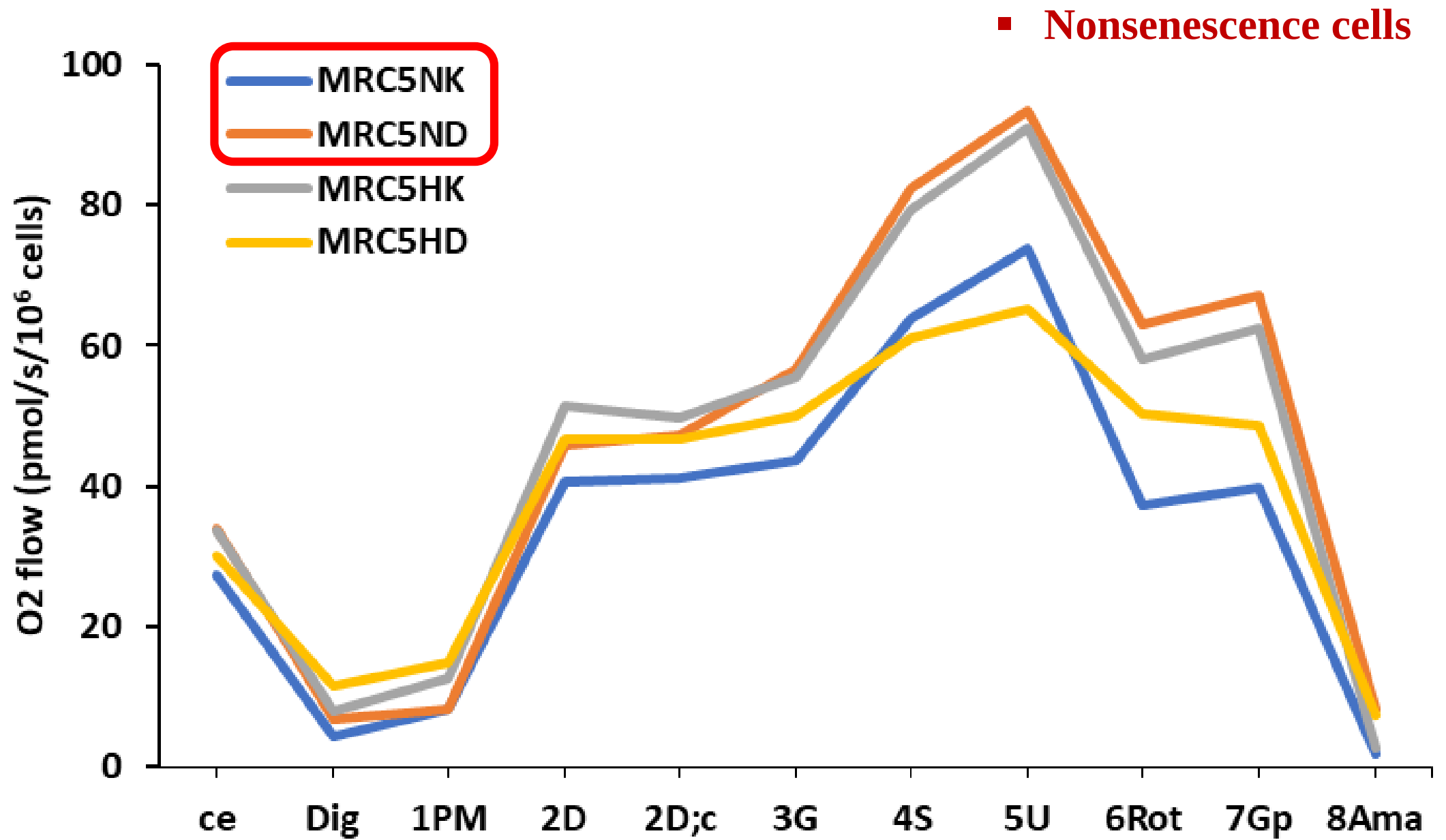


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Mitochondrial respiration	Measurement of respiration

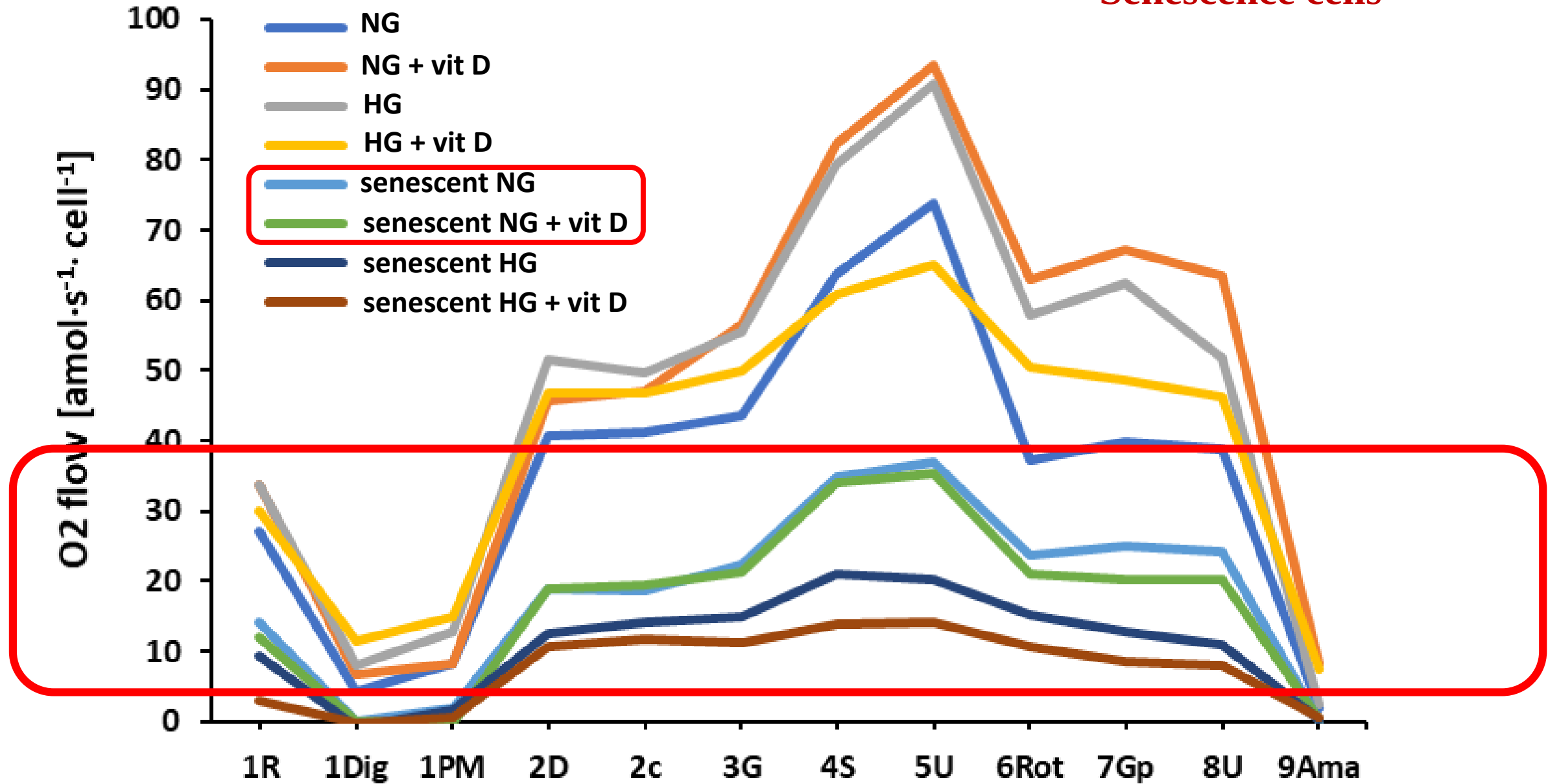
■ Mitochondrial respiration





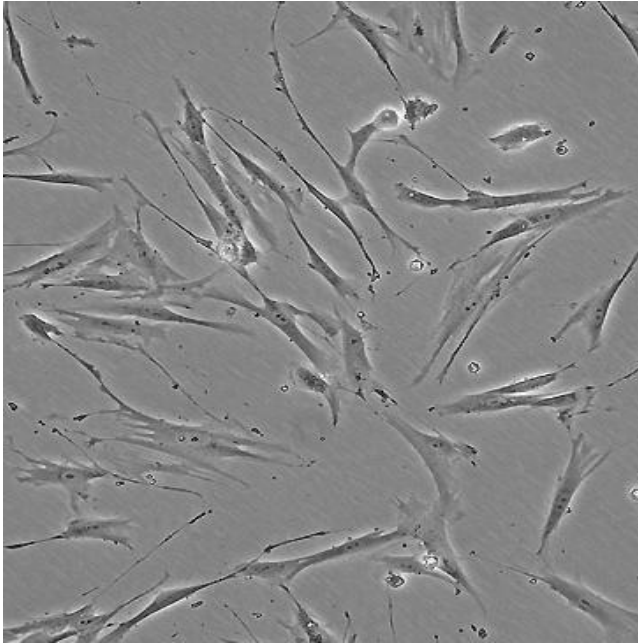


■ Senescence cells



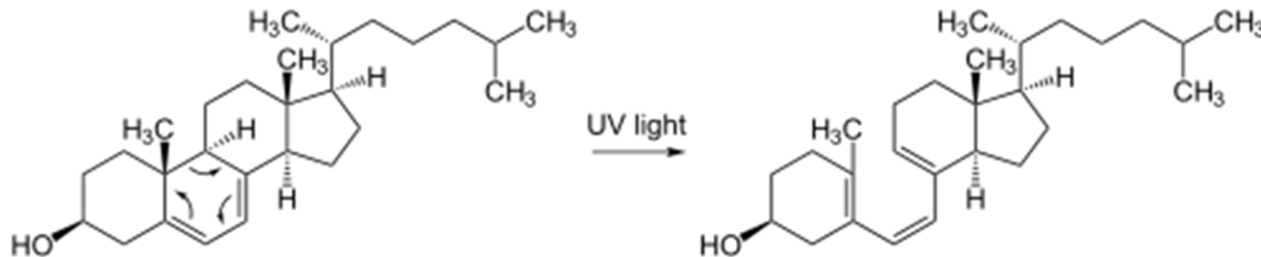
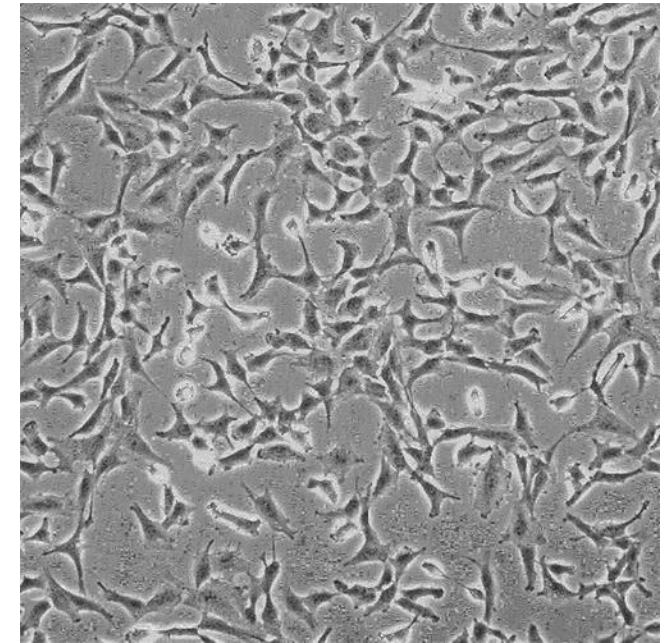
Materials and methods

Human lung cells (**MRC-5**)



- in passage 8 (p8) as normal cells
- in passage 21 (p21) as senescent cells
- senescence induced by 100 μM H_2O_2

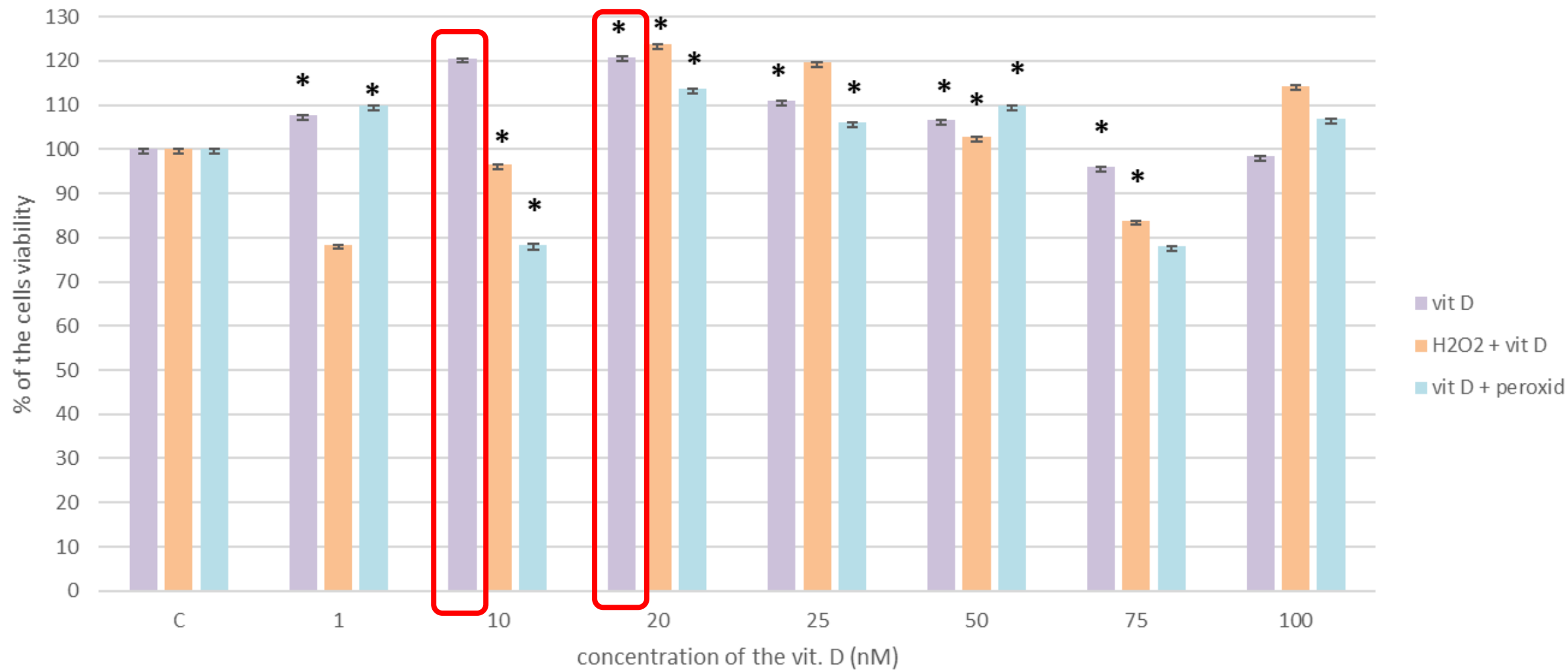
Human astrocytes (**NHA5**)



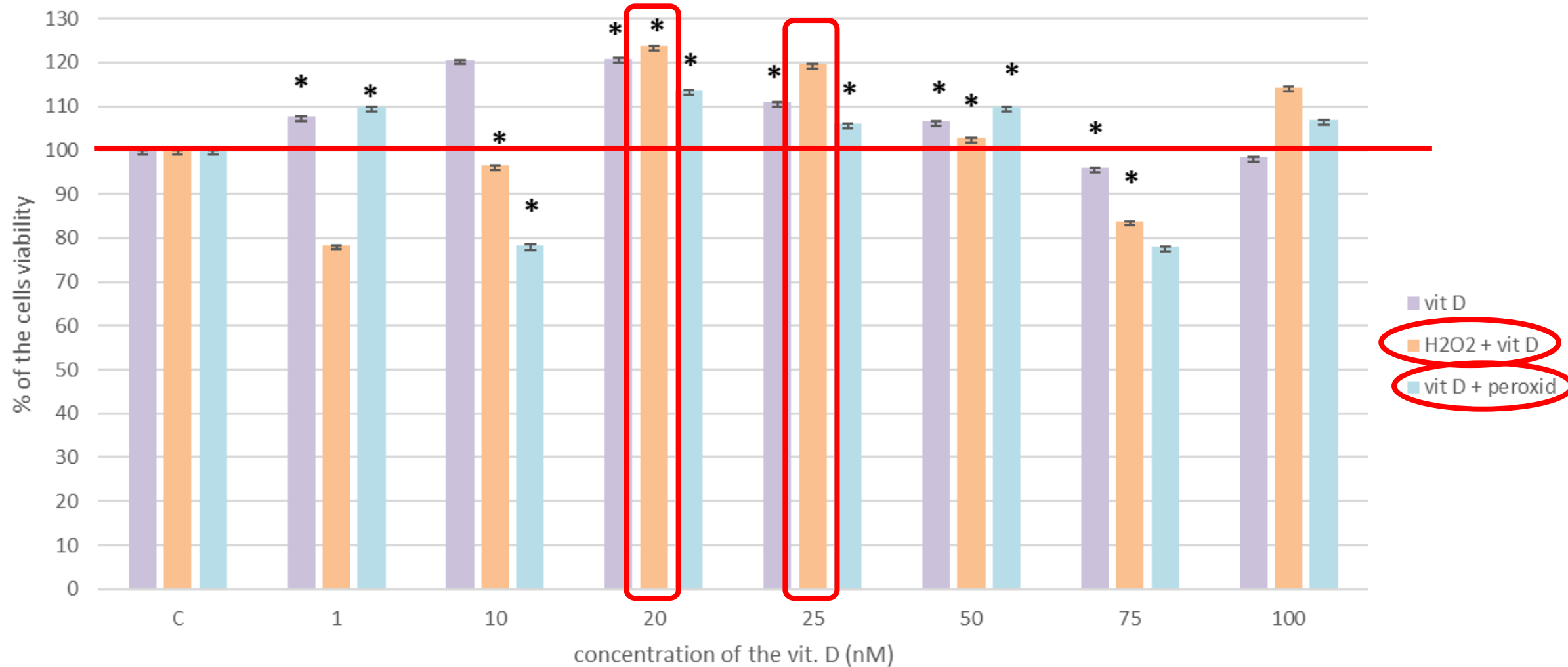
Vitamin D

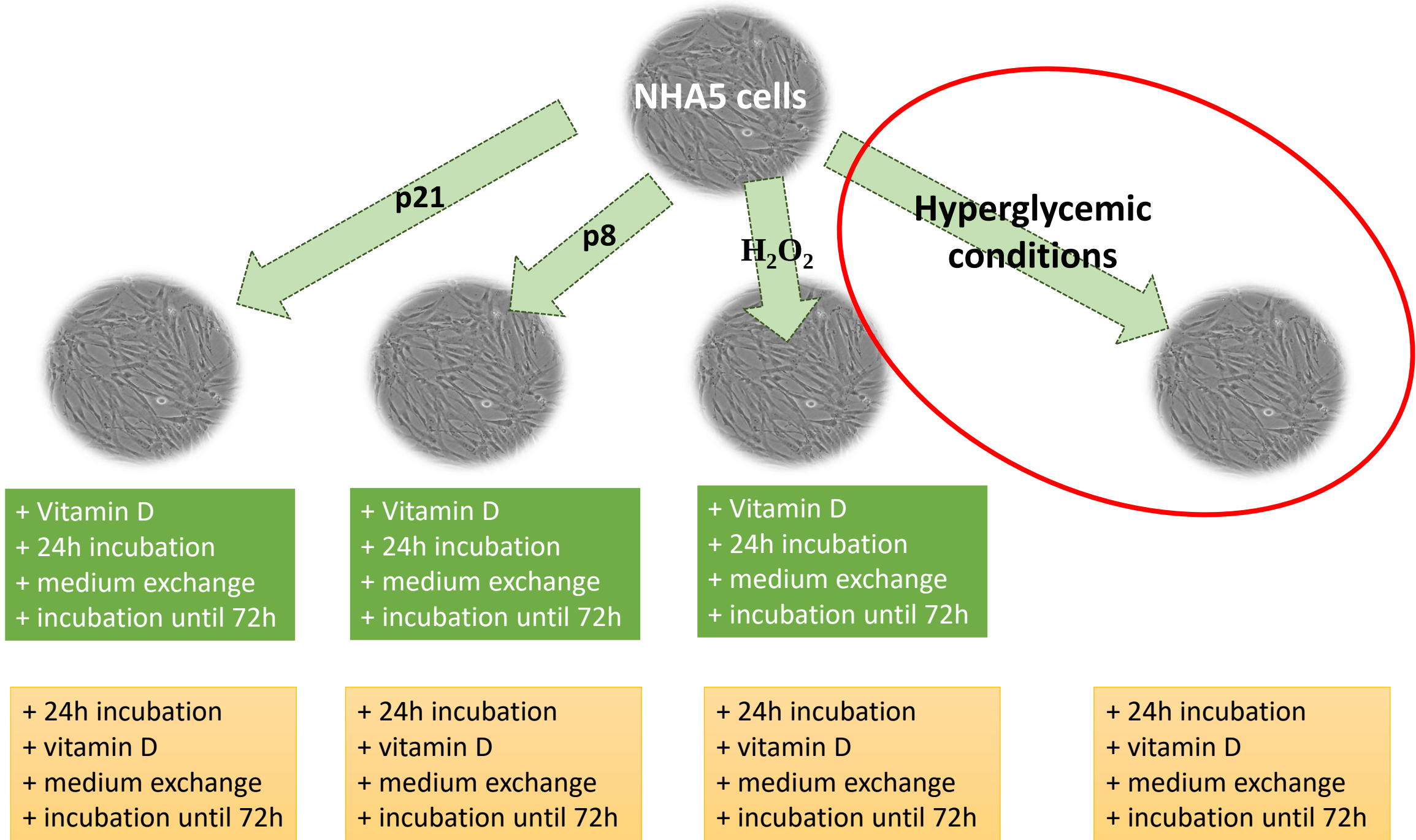
- 1, 10, 20, 25, 50, 100 nM
- in DMSO
- in EtOH

Astrocytes, 24h



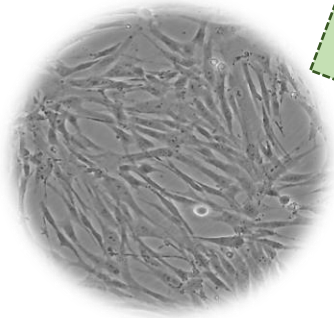
Astrocytes, 24h





NHA5 cells

- + seeded the cells under HG conditions
- + incubated for 24 hours
- + affected cells with vitamin D for 24 hours



Vitamin D

Damaged the cells with H_2O_2 and then affected vitamin D

Added vitamin D to the cells and then damaged them with H_2O_2

not hyperglycemic
NG

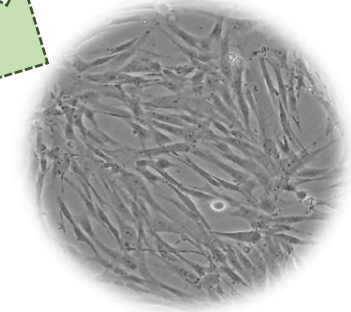


Vitamin D

Damaged the cells with H_2O_2 and then affected vitamin D

Added vitamin D to the cells and then damaged them with H_2O_2

- + seeded the cells under normal conditions
- + incubated for 24 h
- + changed the NG conditions to HG
- + addition of vitamin D

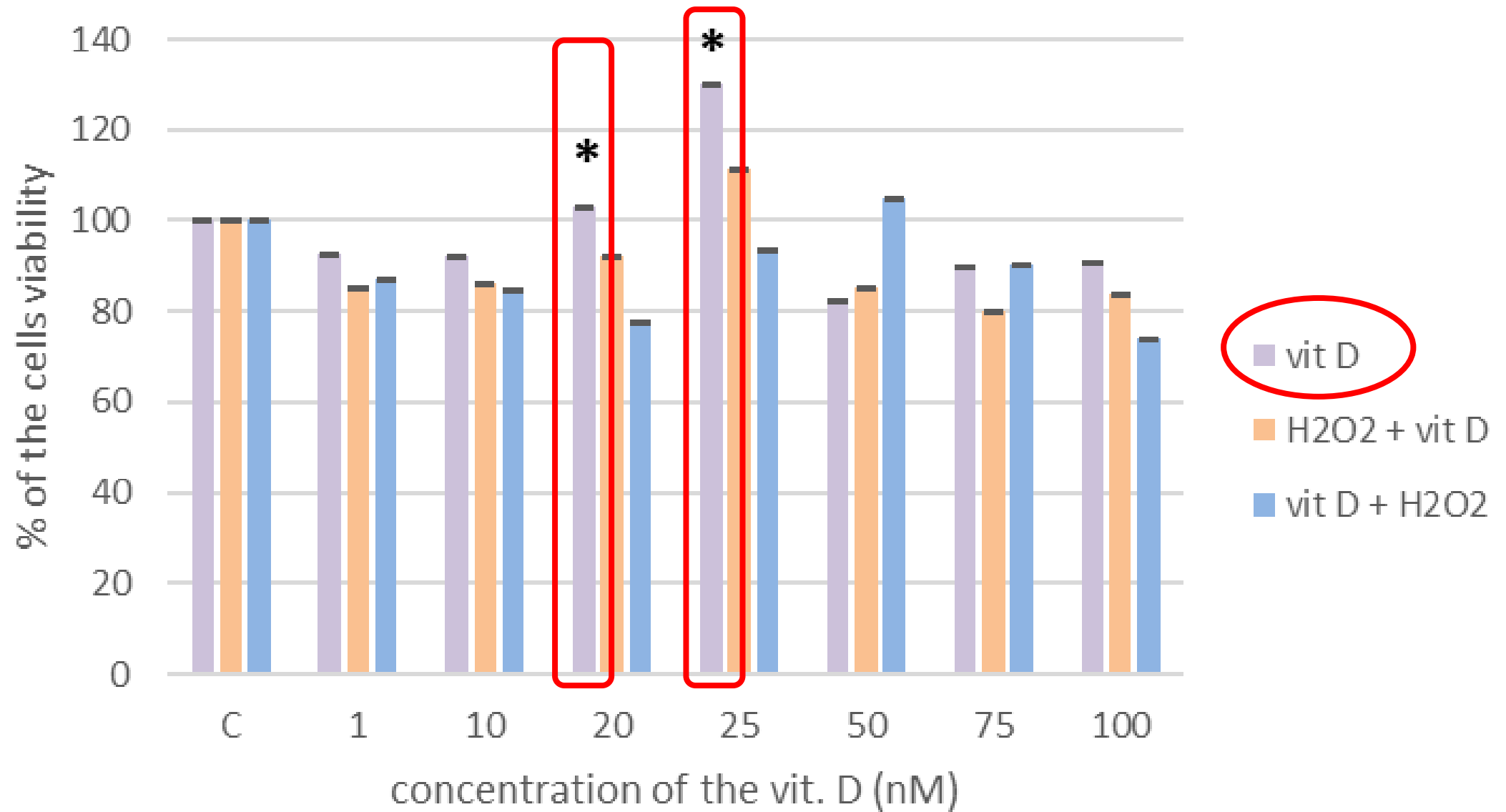


Vitamin D

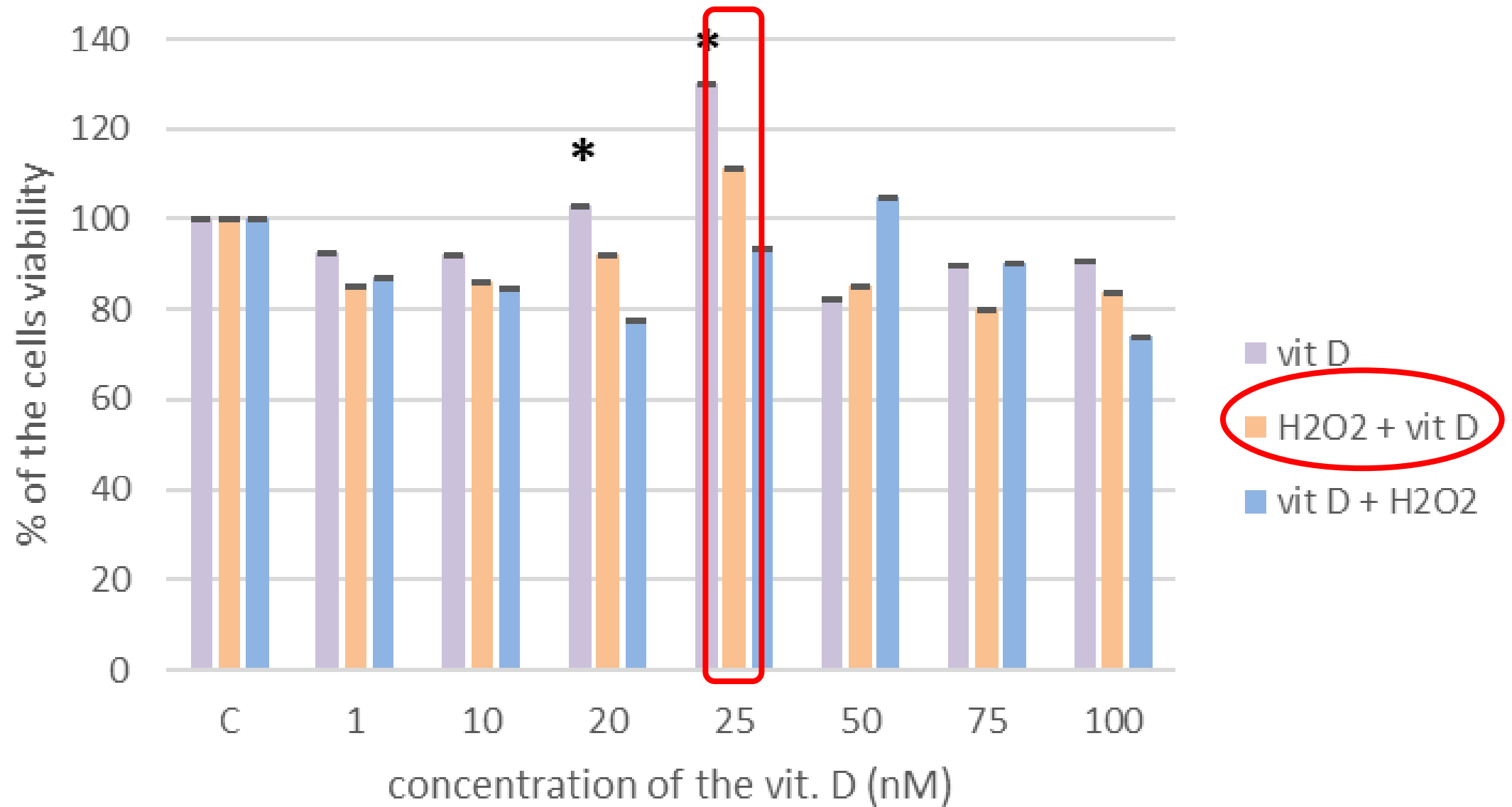
Damaged the cells with H_2O_2 and then affected vitamin D

Added vitamin D to the cells and then damaged them with H_2O_2

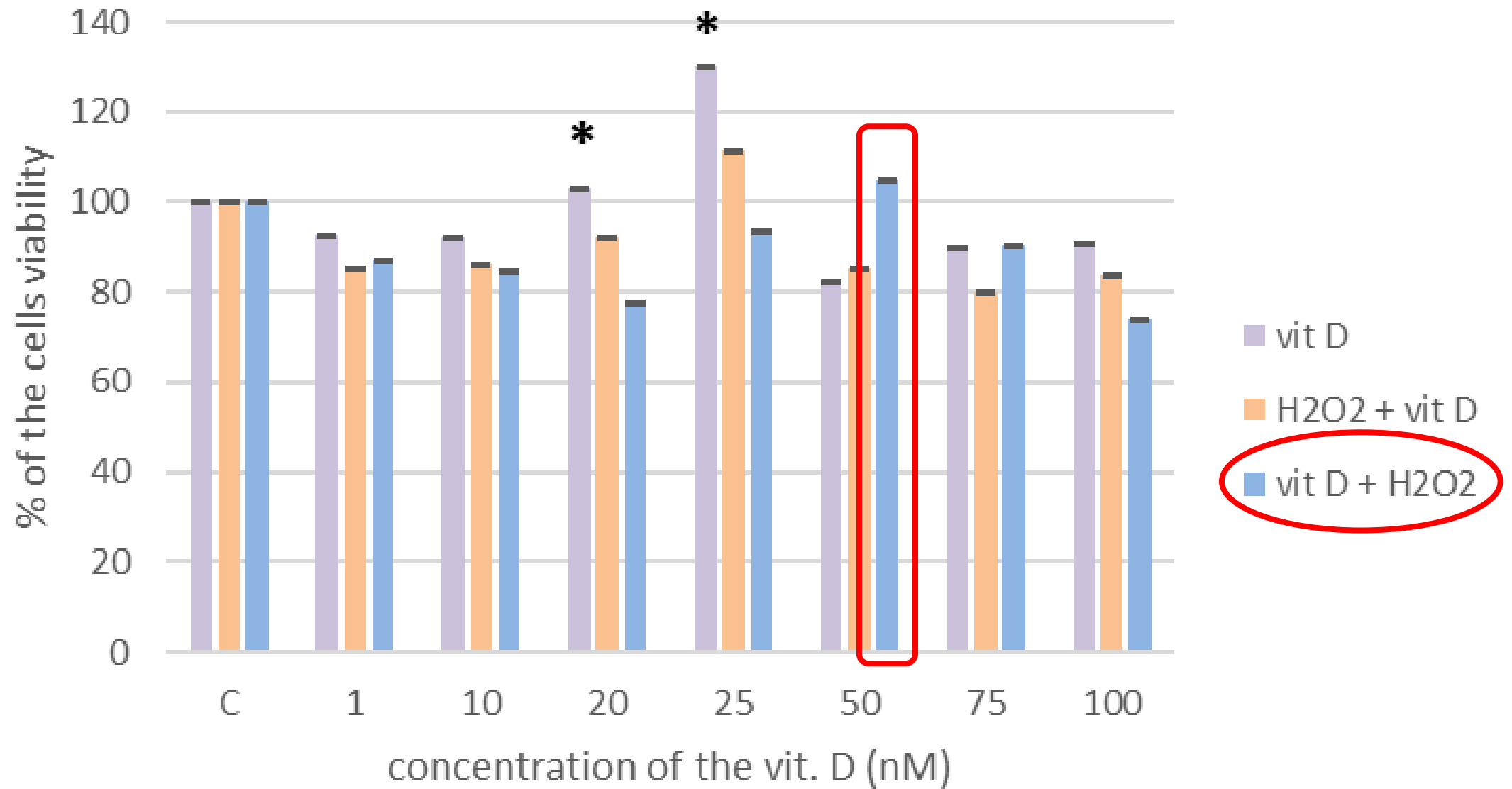
Astrocytes, HG, 24h



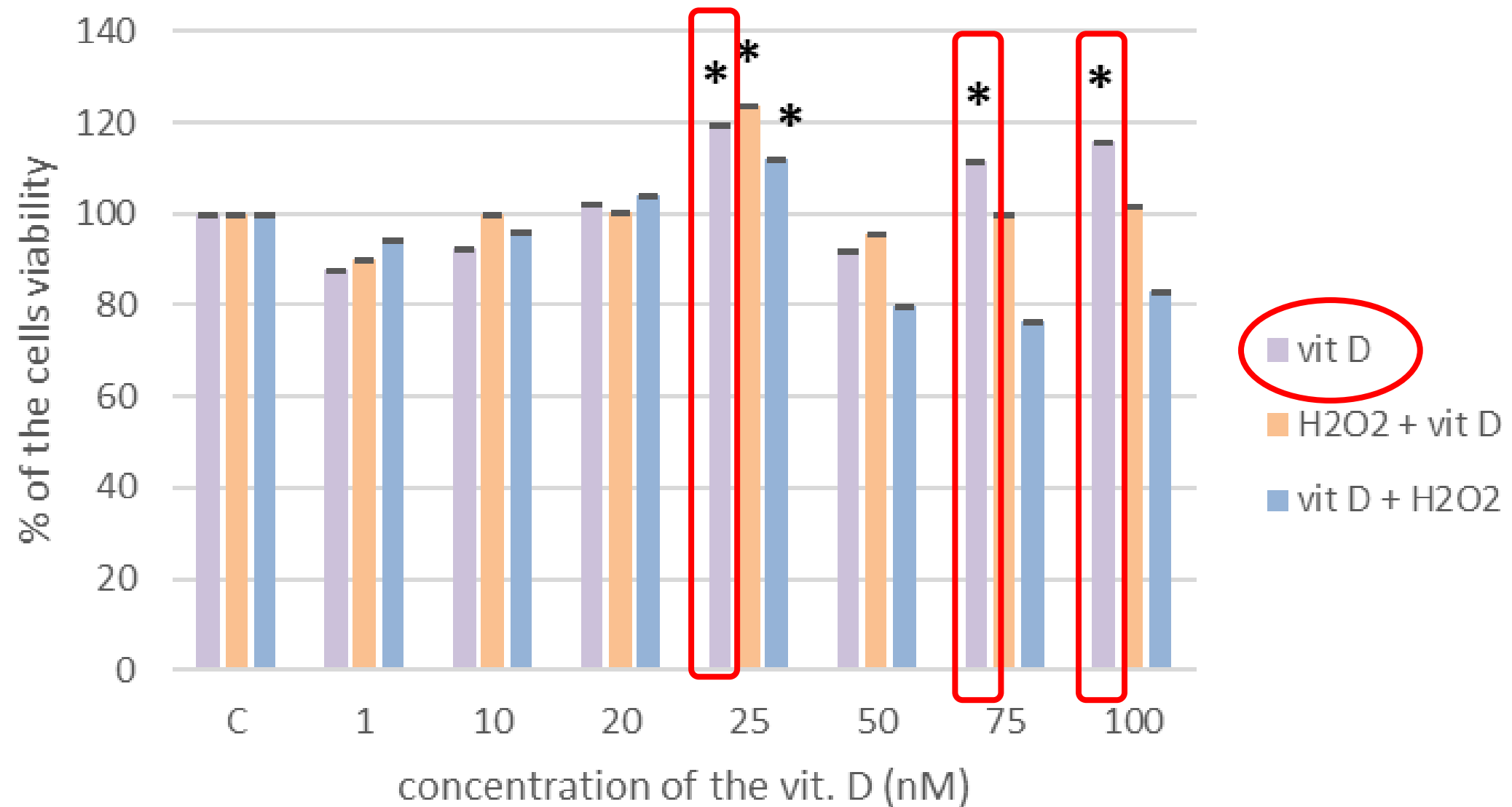
Astrocytes, HG, 24h



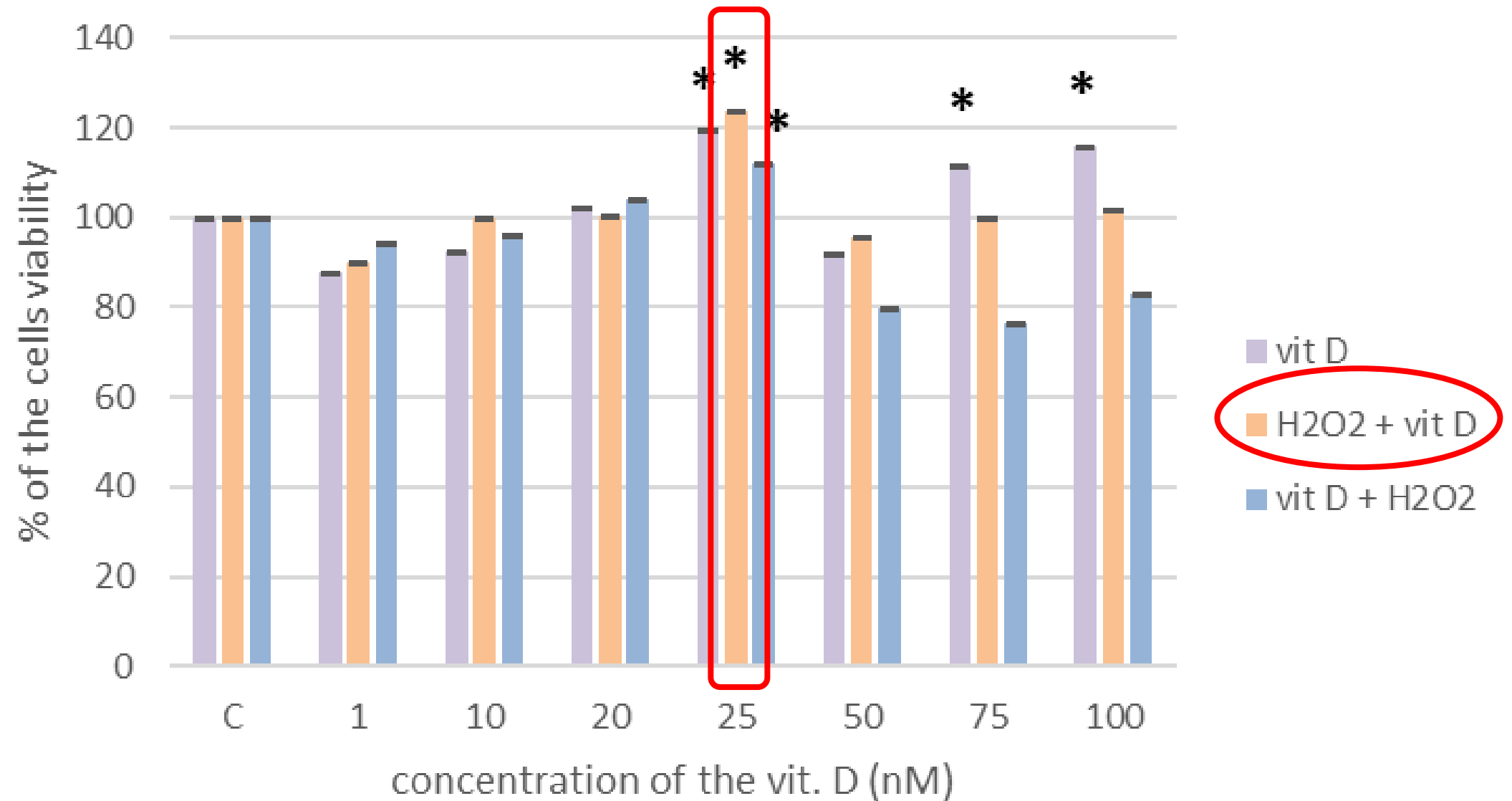
Astrocytes, HG, 24h



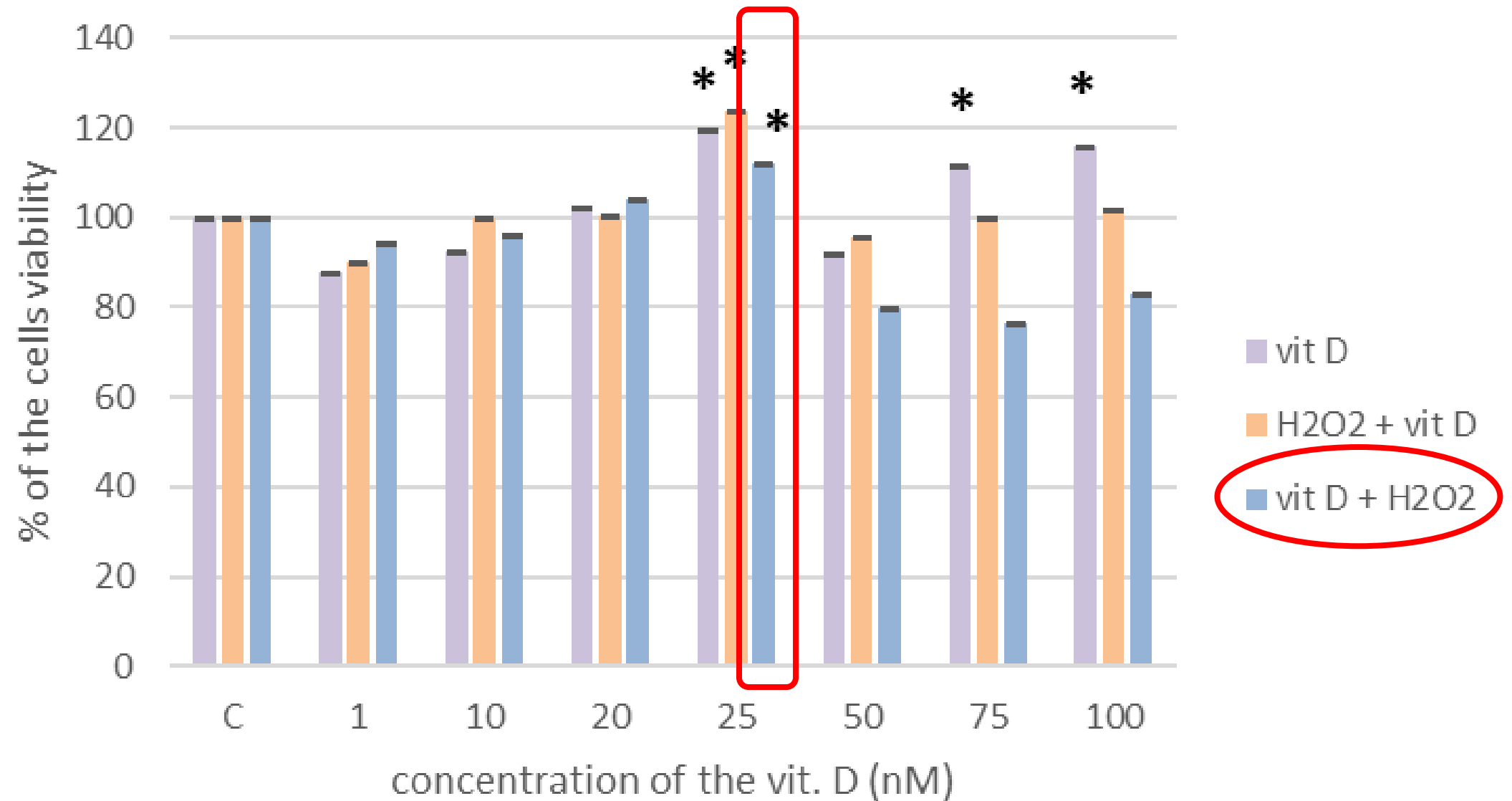
Astrocytes, NG, 24h + HG



Astrocytes, NG, 24h + HG



Astrocytes, NG, 24h + HG



Summary

In senescent MRC -5 cells vitamin D in concentration of 25 nM:

- Positively affected cell viability compared to control cells that were not affected by vitamin D
- We observed a slight decrease of cells in the G1 phase compared to the control and an increase in G2/M phase
- It reduced the expression of the p21 gene at the transcriptional and protein levels

Summary

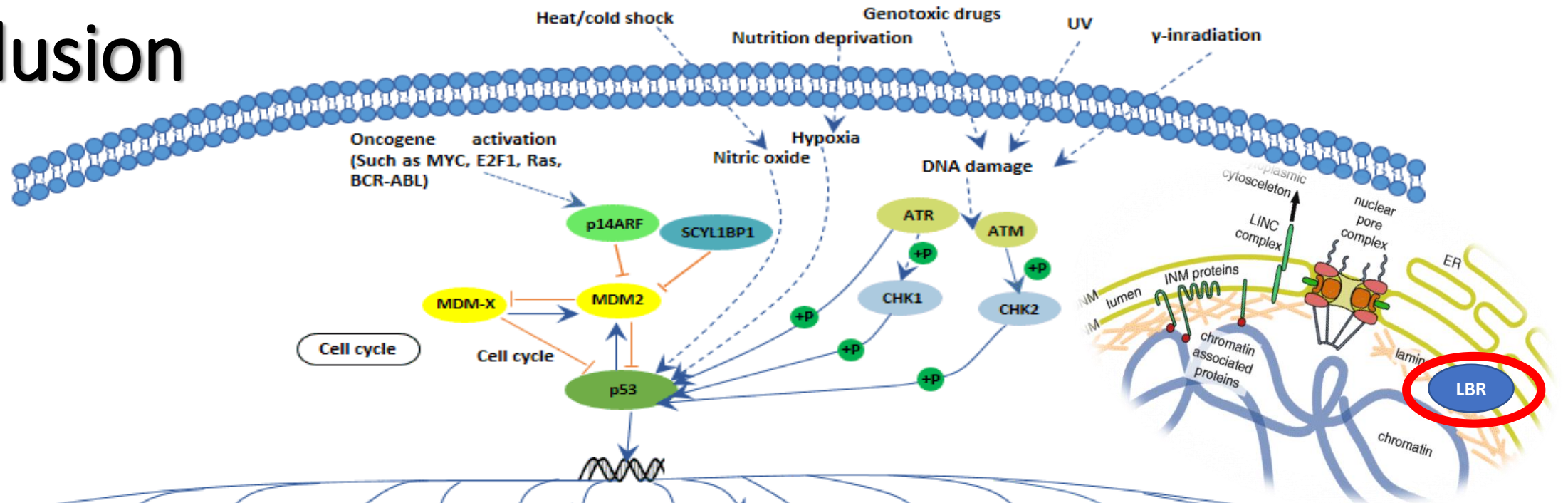
In senescent MRC -5 cells vitamin D in concentration of 25 nM:

- It reduced the expression of the p53 gene at the protein level
- It increased the expression of the LBR gene at the transcriptional level
- Vitamin D reduced LC3 protein levels

In senescence astrocytes vitamin D in concentrations of 25 a 50 nM:

- Positively affected cell viability

Conclusion



In senescent MRC -5 cells vitamin D at concentration of 25 nM improves survival of senescent cells by **inhibiting expression of cell cycle inhibitor** and by **increasing the expression of LBR gene**.

P53 negative feedback

Inhibition of IGF-1/mTOR pathway

Exosome mediated secretion

Apoptosis

Cell cycle arrest

Thank you to my colleagues for their support in carrying out these experiments:

Ingrid Žitňanová, Mária Janubová, Zuzana Sumbalová

Thank you for your attention



References

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